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# Extraction and Separation of Phytoestrogens from Over-the-Counter Supplements

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# **Extraction and Separation of Phytoestrogens from Over-the-Counter Supplements**

Graham Grinnell

Worcester Polytechnic Institute

June 20, 2013

**Abstract**

This study involves the analysis of over-the-counter phytoestrogen supplements due to their anti-proliferative effects on breast cancer cells. Phytoestrogens have been used as alternatives to traditional hormone therapy in postmenopausal women. TLC and HPLC were employed to optimize extraction and separation of these compounds from over-the-counter supplements. Testing of Promensil, Black Cohosh, and Soy Isoflavone supplements revealed that significant anti-proliferative effects observed in Promensil and Black Cohosh may be attributable to their combination of the phytoestrogens daidzein and biochanin A.

## **Acknowledgements**

Sincerest gratitude goes out to all of the following:

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If anyone is missing in the list above, please consider yourselves thoroughly thanked as well. This project would not have succeeded if it weren't for everyone involved generously donating their time and resources.

Thank you!

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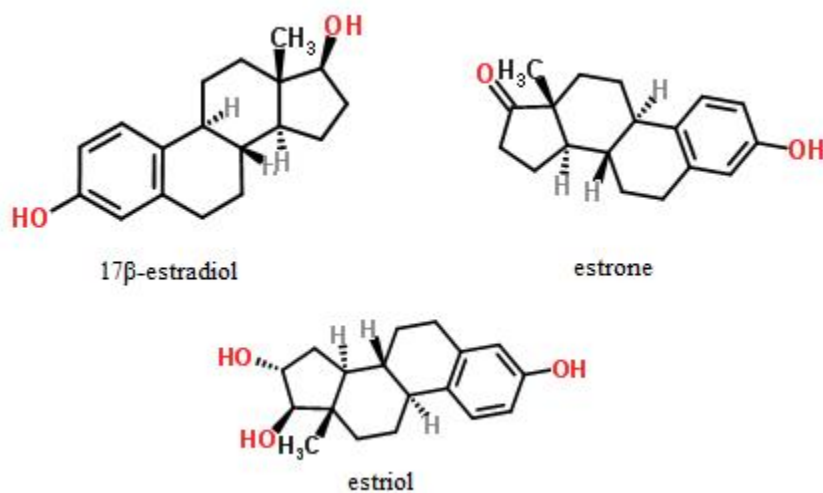
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## **Background**

*This study is part of an ongoing work in the Biology and Biotechnology Project Lab at Worcester Polytechnic Institute. Beginning in 2007 with Caron's thesis, in which it was found that phytoestrogens present in the over-the-counter supplement Promensil have a profound inhibitory effect on the proliferation of MCF7 breast cancer cell lines (Caron, 2007), several successive studies have focused on these steroidal compounds and their effects on breast and prostate cancer cells. To determine what compounds in isoflavone supplements invoke these reactions, it is imperative that they be extracted from the supplements and isolated for further testing. This may be achieved through the use of separatory techniques such as chromatography. The first chapter of this study explores methods of extraction and analyzes them through thin-layer chromatography; the second chapter employs high-performance liquid chromatography to identify major compounds of interest from the separated constituents of extraction mixtures.*

Estrogens are a class of steroidal hormones produced in mammals which promote differentiation of female reproductive organ tissues. They are mainly produced in the ovaries and adrenal glands; however, trace amounts are also produced elsewhere in the body. Three of these estrogens occur naturally within humans:  $17\beta$ -estradiol, estrone, and estriol (Turner, 1971).

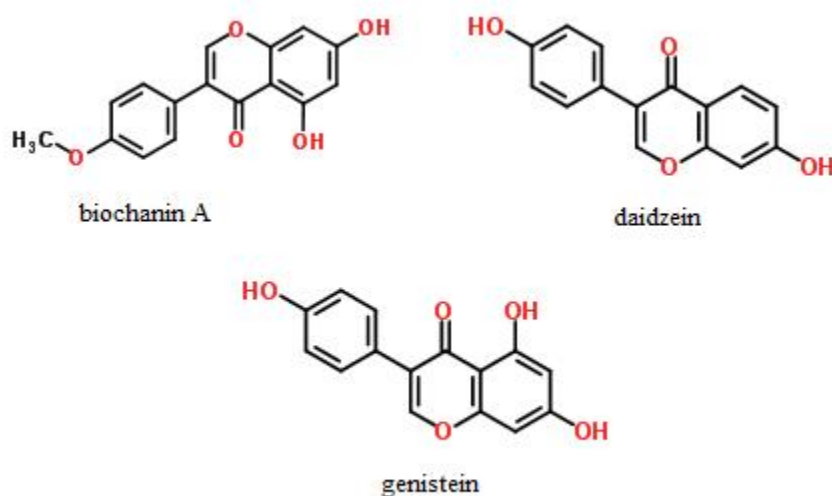


**Figure 1-** The three naturally-occurring human estrogens and their structures (chemspider.com, 2013).

$17\beta$ -estradiol is generally the most active of the three, and may be converted to the less potent estrone or estriol by estrogen-metabolizing enzymes to regulate hormone levels. Other metabolite products of these estrogens may also be present within the body at times (Vaskivuo, 2005). When a female reaches menopause her production of estrogens in the ovaries falls drastically, resulting in symptoms such as menopausal flushes, increased risk of cardiovascular diseases, and a decrease in bone density which may lead to osteoporosis (Hadley, 1988; Hulley,

1998). To relieve these symptoms, hormone replacement therapy may be employed. Unfortunately there are health risks associated with this form of therapy, such as an increased risk of pulmonary embolism and possibly even breast cancer; this being the case, many women have turned to over-the-counter phytoestrogen supplements rather than traditional hormone replacement therapy (Hulley, 1998; David, 2001).

Phytoestrogens (Fig. 2), another name for the isoflavones, are a class of plant-produced steroidal compounds which, in their native organisms, serve the purpose of hormones to induce flowering, growth, and healing of wounds (Turner, 1971). They are most prevalent in the leguminous plants including but not limited to beans, peas, clovers, and soy (Harjo, 2007).



**Figure 2-** Examples of phytoestrogens and their structures (chemspider.com, 2013).

In the human body, however, phytoestrogens have many intriguing effects and health benefits. Their structural similarities to the human estrogens allow them to bind to estrogen receptors and in doing so provide much of the same effect as estrogens (Vacek, 2008). Due to the structural similarities, they exhibit competitive binding with natural estrogens and therefore they are most effective in low-estrogen environments such as those found in postmenopausal women (Messina, 1999). A synthetic isoflavone, ipriflavone, displayed potential for the prevention of osteoporosis by increasing bone mineral density in postmenopausal women (Messina, 1999). In 1993 Coward *et al* observed an overall lower incidence of breast cancer in premenopausal Asian women, which may be linked to diets containing high amounts of soy product rich in phytoestrogens (Coward, 1993). This may be attributable to the antagonistic qualities of phytoestrogens: estrogen-receptor positive breast cancer would be inhibited by their competitive binding with estrogens (Zajchowski, 1993). These beneficial effects have led to a large market for over-the-counter phytoestrogen supplements, but the possibility of these compounds acting as estrogen agonists when in the body leads one to believe they may have the same downsides and risks as



traditional hormone replacement therapy, especially since they are unregulated from lot to lot (Setchell, 2001).

## CHAPTER I: Supplement Extraction and Thin-Layer Chromatography

### Introduction

The extraction and isolation of phytoestrogens has been studied for well over a century: a study by Perkin and Newbury in 1899 was the first to isolate genistein from the legume *Genista tinctoria*, from which the name of the compound was derived (Perkin, 1899). Their experiment involved an extraction first in acid, then an alcohol which is assumed to be methanol: they do not clarify in their writing. While it was primarily a study of the coloring compounds present in the legume, this inadvertent discovery of a new compound would facilitate interest in the further extraction and categorization of phytoestrogens. In 1941 Walter experimented on soybeans to determine further physical and chemical properties of genistein while also refining the isolation procedure (Walter, 1941). The solvent of choice remained methanol, but experimentation with different solvents would become an important part of the phytoestrogen extraction process.

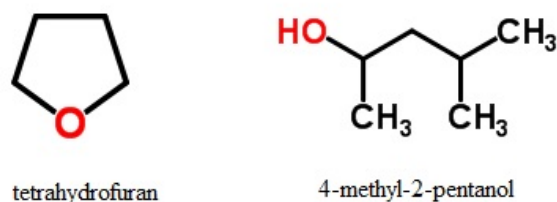
Farmakalidis and Murphy found that both genistein and daidzein could be extracted from soyflakes through the use of acetone and isolated from the extract through silica gel column chromatography using an elution mixture of 9:1 chloroform and methanol (Farmakalidis, 1985). Murphy later tested acetonitrile, acetone, ethanol, and methanol extractions of soy and found acetonitrile to be the most efficient for extracting phytoestrogens (Murphy, 2002). Other experiments have showed that hexane, 2-propanol, isopropanol, and other relatively plentiful organic solvents were viable alternatives to methanol or ethanol extractions (Vacek, 2008).

Setchell *et al* used methanol in an attempt to extract as much phytoestrogen content as possible from 33 varying over-the-counter supplements. Since these supplements are undefined, unregulated mixtures of plants, the group's intent was to verify how accurate the producers' claims were regarding their products' phytoestrogen content and therefore an all-inclusive method of extraction was necessary (Setchell, 2001). Caron later used the same method in extracting genistein from the over-the-counter product Promensil (Natrol); the resulting methanol extract was observed to have anti-proliferative effects on MCF7 breast cancer cell lines (Caron, 2007).

As a result, Promensil, as well as two other over-the-counter supplements, Soy Isoflavones (Natrol) derived from soy plants and Black Cohosh (Solaray) derived from the black cohosh plant *Cimicifuga racemosa*, will be the focus of this study. The Promensil packaging states that one dose contains 40 mg of isoflavones without indicating any compounds in particular. Natrol claims its Soy Isoflavones contains 20 mg of genistein, 17.5 mg of daidzein, 6.25 mg glycitein, and 6.25 mg "other isoflavones" per dose. Solaray's Black Cohosh packaging does not list any specific phytoestrogen content, simply stating "Black Cohosh is known for its phytoestrogen properties." The high variability in the nature of these plant products means

phytoestrogen content may differ significantly between batches if the source plants are not kept in highly controlled environments. Even then, variables such as the seasons may factor in to phytoestrogen content from batch to batch.

Particular phytoestrogens or supplements may require more efficient extraction media than others. To provide a more optimal and efficient extraction method for the three supplements in question, various organic solvents were tested for their extraction efficiency relative to methanol; this would also determine viable alternatives for different situations. Besides methanol, tetrahydrofuran and 4-methyl-2-pentanol (Fig. 3) were selected for extraction testing. THF was selected for its significant polar properties which would better facilitate extraction of compounds such as phytoestrogens. Conversely, 4-methyl-2-pentanol was selected in the interest of seeing whether side chains on a solvent compound might inhibit extraction efficiency.



**Figure 3-** Selected alternative extraction solvents and their structures (chemspider.com, 2013).

## **Materials and Methods**

### **Extraction of phytoestrogens from supplements:**

Supplement extraction was performed according to Caron's method (Caron, 2007). Extraction media were prepared as follows: 80% methanol (V/V in H<sub>2</sub>O), 80% tetrahydrofuran (V/V in H<sub>2</sub>O), and pure 4-methyl-2-pentanol. Three different over-the-counter supplements were chosen for extractions: Promensil (Natrol), Black Cohosh (Solaray) supplements, and Soy Isoflavones (Natrol) supplements. For each extraction, four tablets of the corresponding supplement were ground by mortar and pestle before being added to 80 mL extraction medium. The solutions were then refluxed with a water-jacketed condenser for 1 hour before being filtered and stored at 4°C. An 80% 4-methyl-2-pentanol (V/V in H<sub>2</sub>O) solution was also prepared to determine whether isoflavones would extract more readily to the organic or aqueous phases. This was only tested with Promensil.

### **Determination of proper spot volumes and plate composition for TLC:**

This initial test used two alumina TLC plates running concurrently. Methanol extractions of each of the three supplements were tested; a micropipettor was used to spot one plate with 20 µL and the second plate with 10 µL of each extraction. TLC was then run for 90 minutes using an 8:3:1 chloroform/methanol/acetonitrile mobile phase as detailed in Caron's experiment. A second test was run to analyze the viability of silica with gypsum versus alumina plates, as well as test smaller load sizes. One alumina plate and one silica with gypsum plate were loaded with the methanol extractions and run using the solvent described above; one sample was first centrifuged at 3000 RPM for 5 minutes and the supernatant was used for its spot. Load sizes were 10 µL, 5 µL, 3 µL, and 1 µL of the extract before centrifugation and 5 µL of the centrifuged sample. All plates were allowed to dry, then analyzed under short-wave UV light. Visible spots were marked for identification by retention factor ( $R_f$ ) values.

### **Determination of TLC clarity among all extracts:**

TLC tests were then run using 5 µL loads of all extraction mixtures on silica with gypsum plates. Again, the samples were centrifuged and the supernatant used for spotting. The 8:3:1 chloroform/methanol/acetonitrile mobile phase was used again and the assay was run for 90 minutes to allow the solvent front to move near the top of the plate. The plates were then allowed to dry. Plates were again visualized and marked under short-wave UV. Two further test plates were run to confirm the system's consistency.

**Identification of phytoestrogens in supplements by TLC:**

Pure biochanin A (Alfa Aesar), daidzein (Enzo Life Sciences), and genistein (Enzo Life Sciences) were obtained for use as pure phytoestrogen standards. These were used to prepare 10 mL solutions of each in 20  $\mu$ M concentrations in methanol. The three standard solutions were then run concurrently with all extraction mixtures. All extracts were again centrifuged before loading the supernatants; the standards were not. Again, 5  $\mu$ L spots were used and TLC was run for 90 minutes with the 8:3:1 chloroform/methanol/acetonitrile mobile phase. The plates were analyzed and marked under short-wave UV.

**Testing the TLC assay using silica F<sub>254</sub> plates:**

Caron's TLC experiment used silica F<sub>254</sub> plates; for consistency and to improve clarity of the TLC assay, these were chosen to be used for further testing rather than silica with gypsum plates. One test plate was run with all standards and extracts. A spot test was employed to ensure the standards could be observed on the silica F<sub>254</sub> plates using short-wave UV. Both centrifuged and non-centrifuged samples were spotted.

**Determination of a solvent system to improve TLC clarity:**

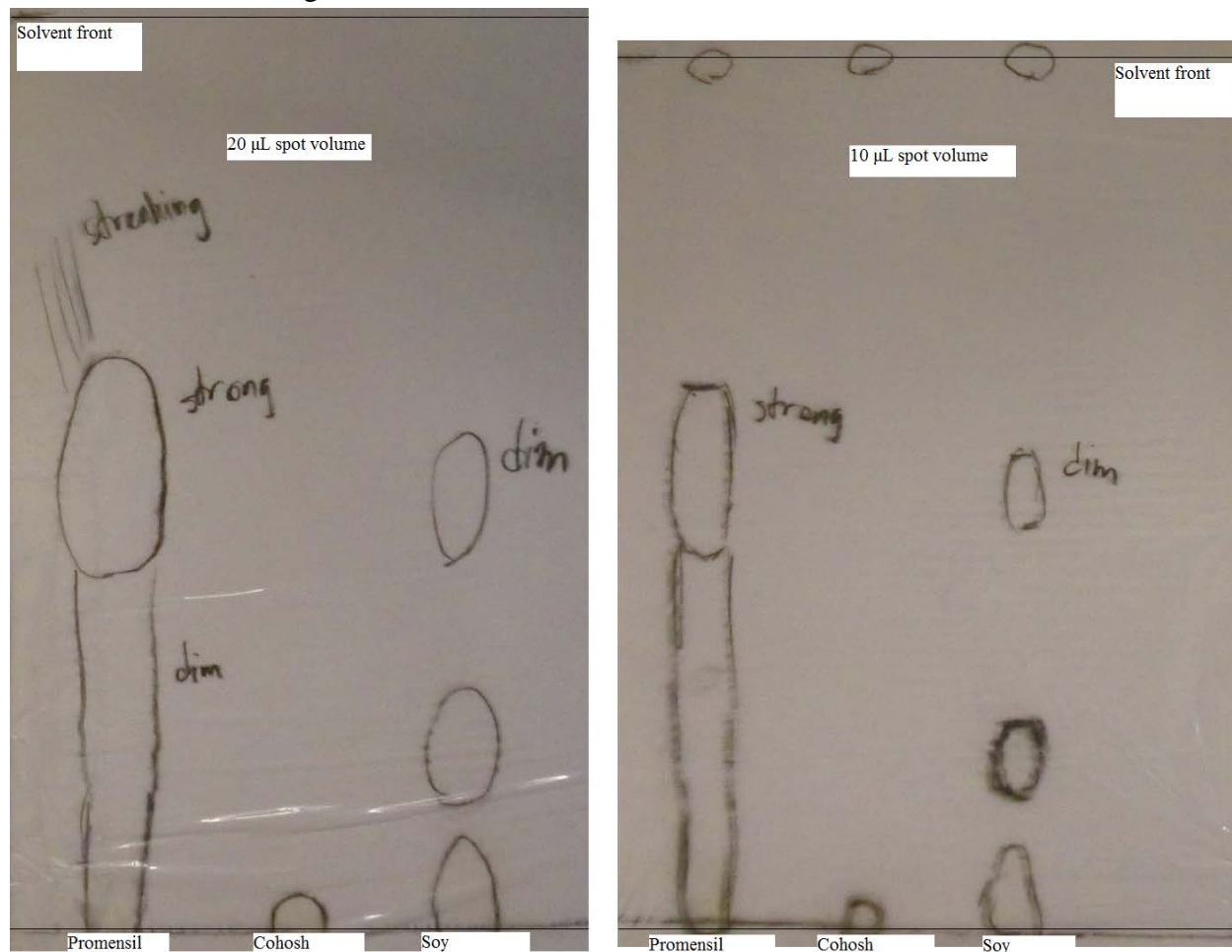
After the spot test, it was observed that the standards were in fact present on the TLC plates, but they were along the solvent front. In an attempt to remedy this situation several small-scale solvent system tests were run to determine a more optimal ratio of solvents. The amount of chloroform was increased due to it having the lowest elutropic value (Hofmann, 1977) of the three solvents, then acetonitrile was also increased to effectively decrease the overall amount of methanol in the system. 12:3:1 and 12:3:2 mixtures were tested. Another small-scale test was used with 12:3:1, 12:2:1, and 12:1:1 ratios on silica with gypsum to test the effect of varying amounts of methanol, which had the highest elutropic value of the three solvents (Hofmann, 1977).

**Identification of phytoestrogens present in extracts:**

Two TLC plates were run using the 12:2:1 chloroform/methanol/acetonitrile mobile phase identified in the previous test. The plates were again run for 90 minutes, then allowed to dry before being visualized under short-wave UV. R<sub>f</sub> values for all bands were calculated to confirm consistency.

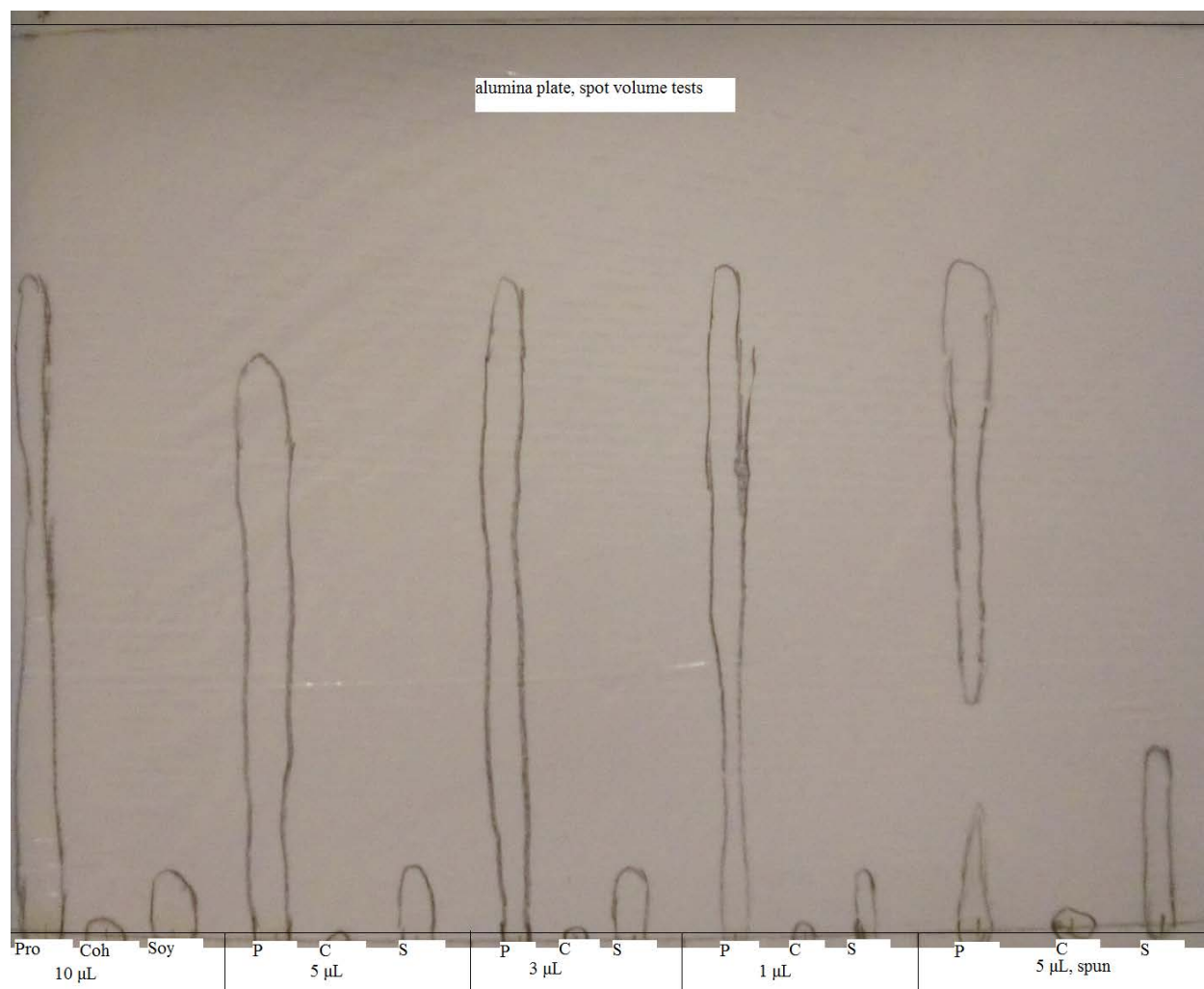
## Results and Discussion

Initial tests were run using Caron's 8:3:1 chloroform/methanol/acetonitrile solvent mixture. Alumina plates were the first available and were used to determine a proper load size (spot volume) for the TLC assay. Two plates were run concurrently: one with 10  $\mu\text{L}$  spots and the second with 20  $\mu\text{L}$ . The resultant plates (Fig. 4) displayed large smudges, suggesting the load sizes were much too large.

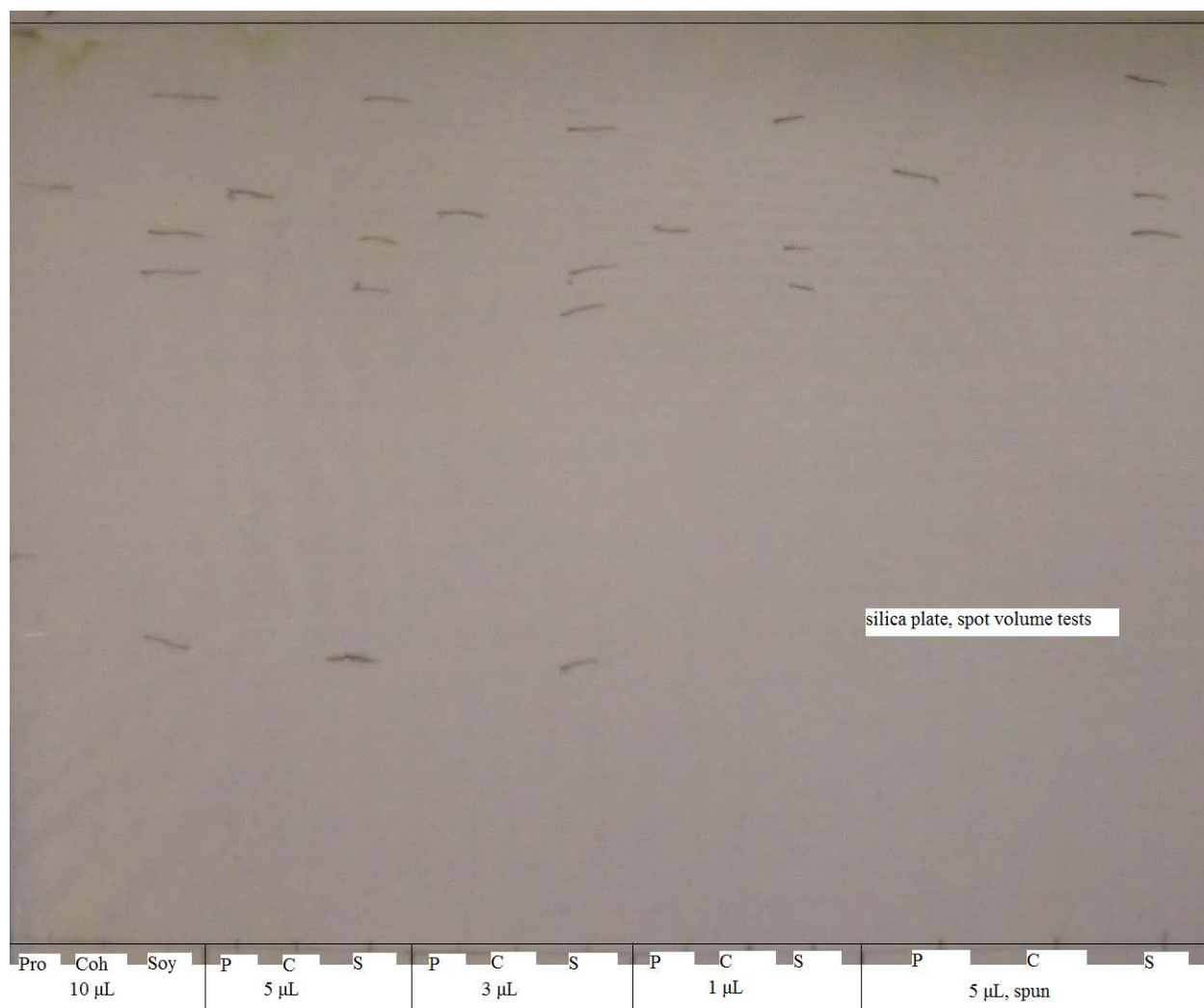


**Figure 4-** Initial spot volume tests using alumina plates. Load volumes of 20  $\mu\text{L}$  (left plate) and 10  $\mu\text{L}$  (right plate) were tested for visibility.

To find a proper load size and test the viability of alumina plates versus silica with gypsum plates, the next test was run with one alumina plate and one silica with gypsum plate concurrently. Results displayed higher clarity and better resolution on the silica with gypsum plate (Fig. 6), on which there were evident bands. The alumina plate (Fig. 5) again had smudging in every lane. It was determined from there that the silica with gypsum plates would be used in later assays and that samples would be spun down before spotting them in 5  $\mu\text{L}$  volumes.



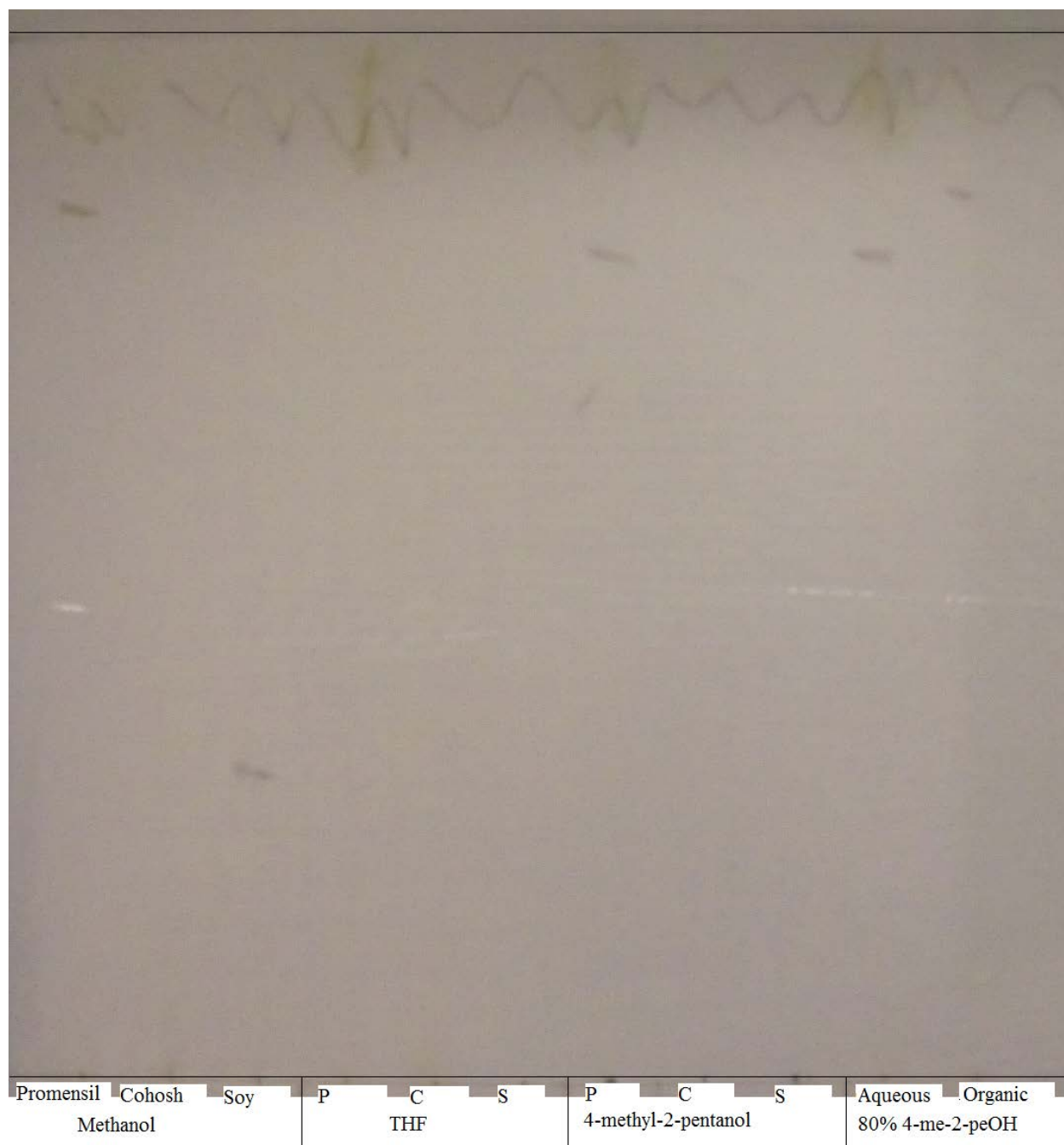
**Figure 5-***Further spot volume tests performed on an alumina plate. Load volumes were 10, 5, 3, and 1  $\mu\text{L}$ ; the rightmost lane was loaded with 5 $\mu\text{L}$  supernatant from a centrifuged sample.*



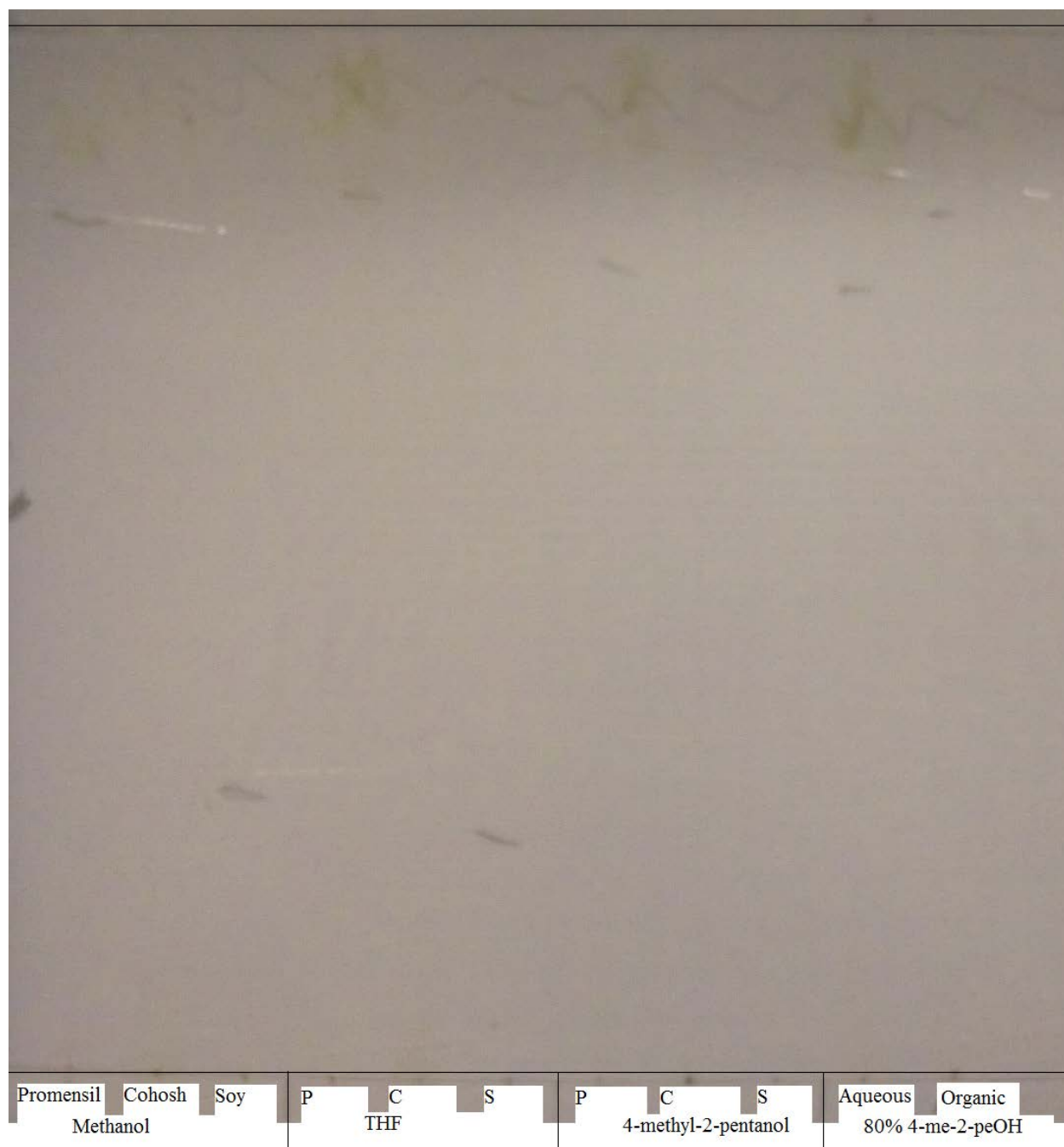
**Figure 6-** The same spot volume tests as Fig. 5 on a silica with gypsum TLC plate.

Two test plates were then run (Figs. 7 and 8) to see how all the extracts would resolve using the determined methods. Bands were observed, yet those in the methanol extraction lanes were inconsistent with those seen in the preliminary tests. To eliminate variables the TLC chamber was cleaned and a fresh solvent mixture was made. A third test was run and excessive smudging was observed (Fig. 9): it was later determined the chloroform used in the second mixture of solvent was contaminated.

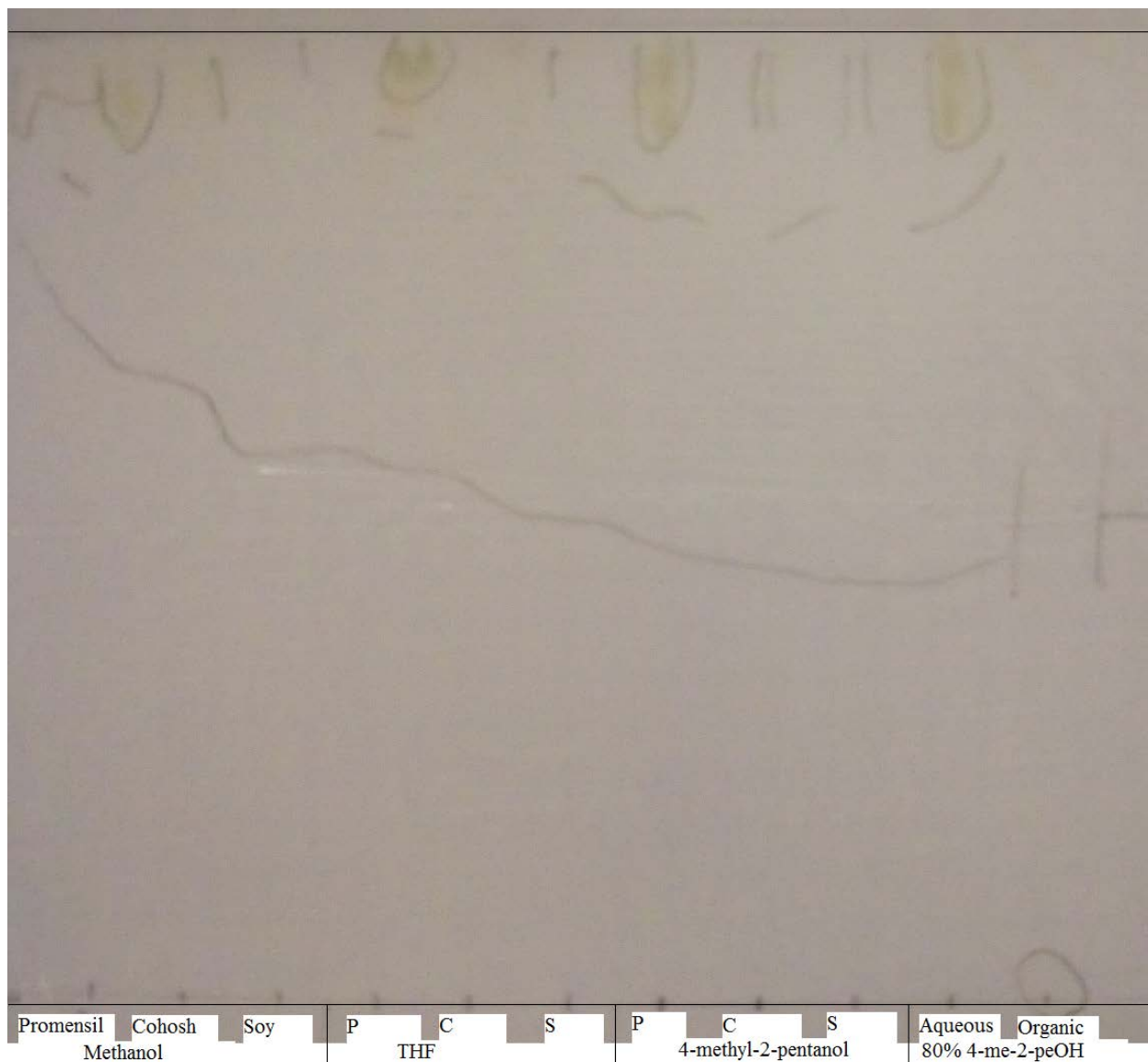




**Figure 7-** Extract test plate #1. THF = tetrahydrofuran. Aqueous and Organic refer to the respective phases in the 80% 4-methyl-2-pentanol extraction.

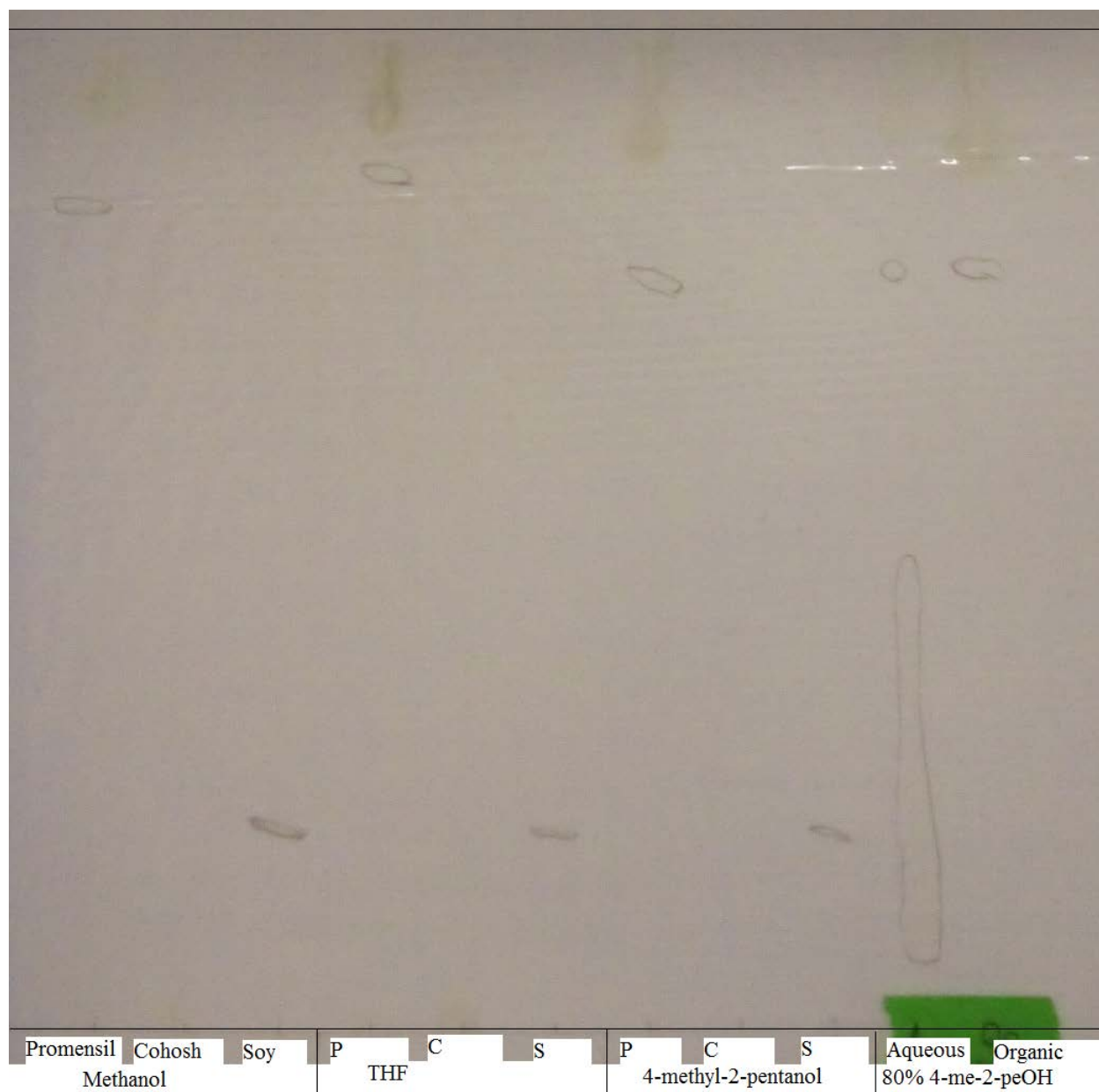


**Figure 8-** Extract test plate #2, run identically to Fig. 7.

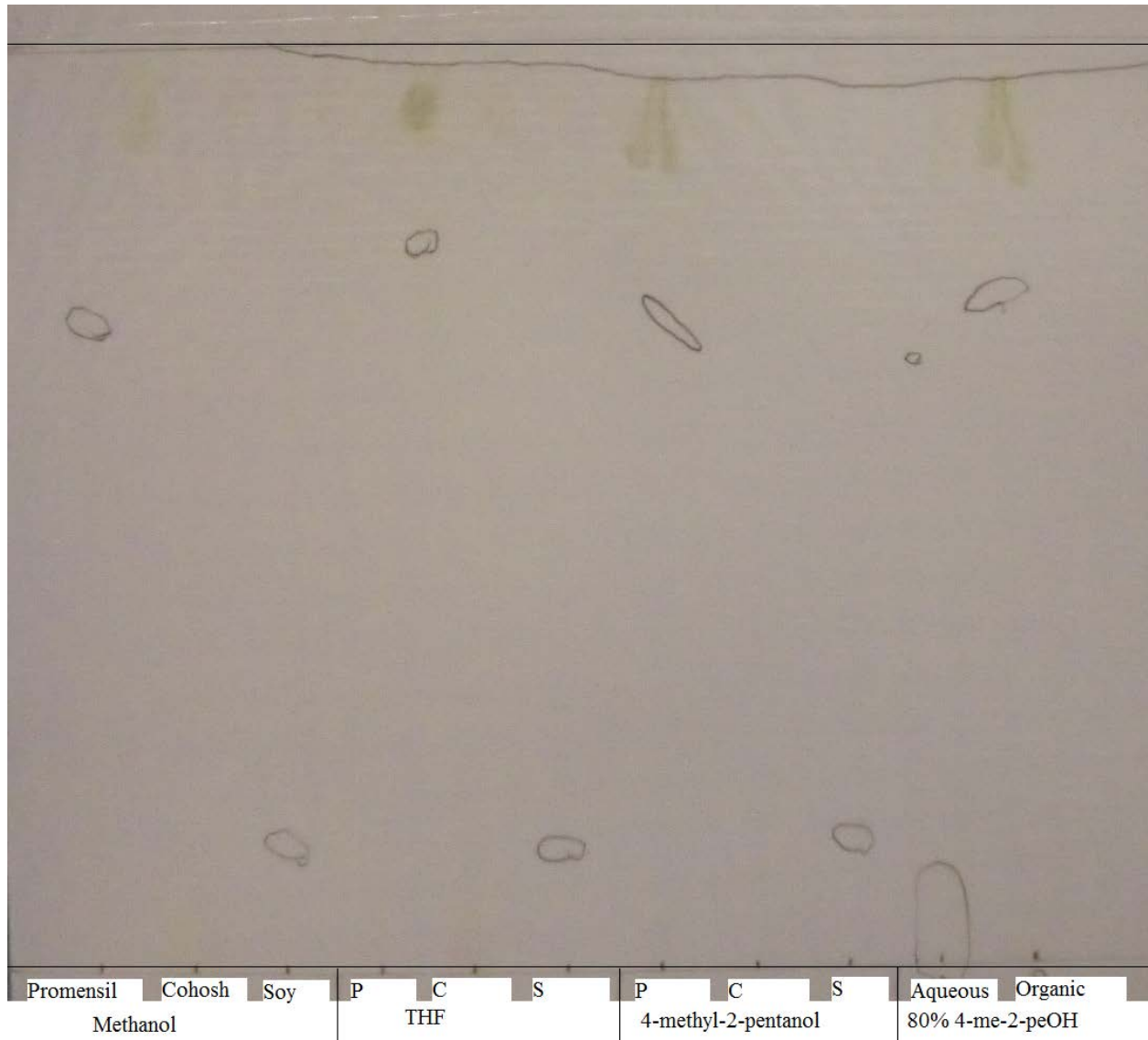


**Figure 9-** Extract test plate #3, run identically to Figs. 7 and 8 but with fresh solvent. Excessive smudging was attributed to contaminated chloroform used in the new solvent.

After obtaining more chloroform for the solvent the test plates were run again; this time, they exhibited relatively clear, albeit few, bands (Figs. 10 and 11). Consistency was also observed visually between the two plates, including the smear at the beginning of the 4-methyl-2-pentanol aqueous phase lane. It was then assumed the TLC assay methodology was working correctly and phytoestrogen standards could be introduced.

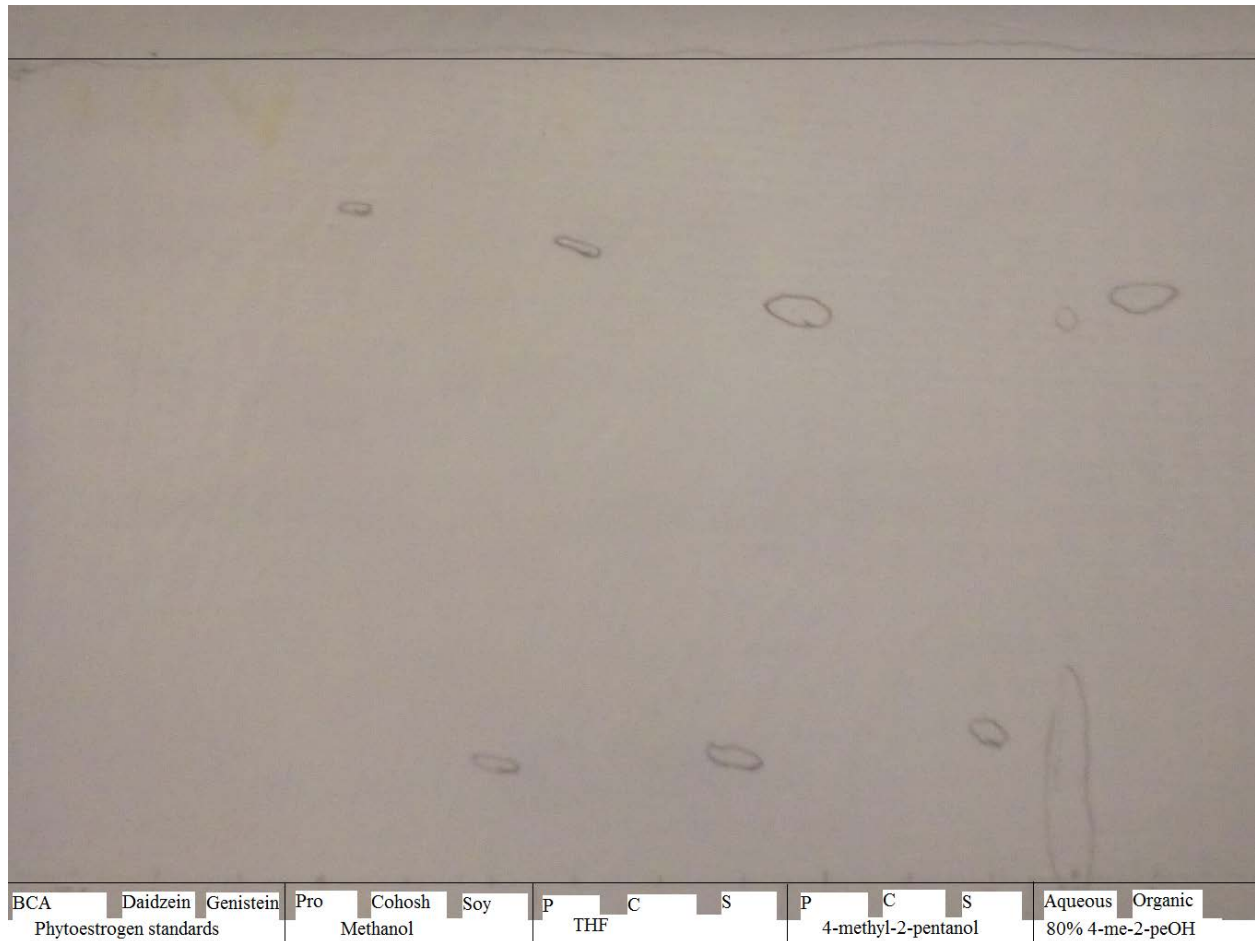


**Figure 10-** Test plate #1 to determine consistency of the TLC methodology.



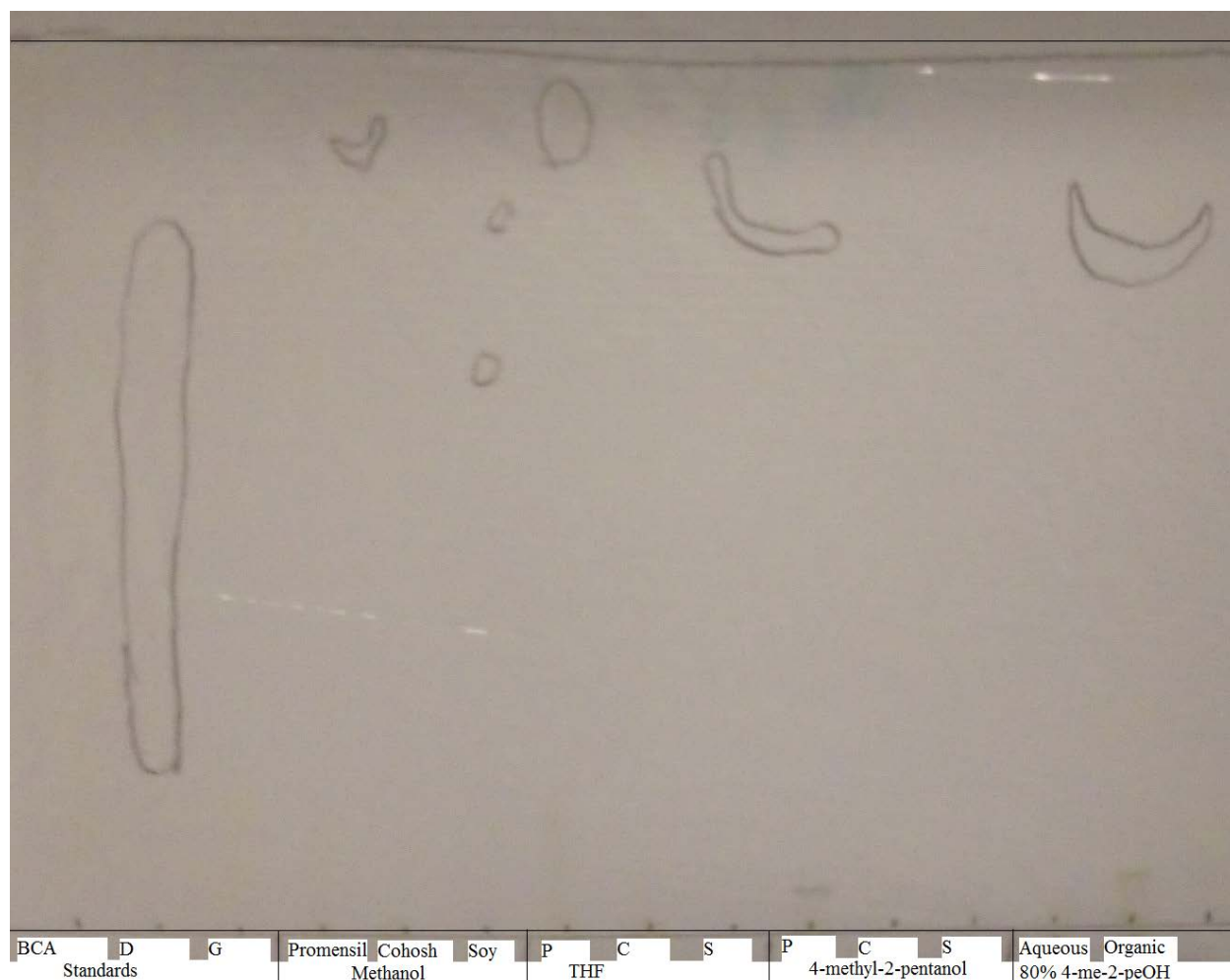
**Figure 11-** Test plate #2 to determine consistency of the TLC methodology. Visualized spots were comparable to those seen in Fig. 10, suggesting consistency in the method.

Three pure phytoestrogen standards were prepared in 20  $\mu$ M concentrations in methanol: biochanin A, daidzein, and genistein. These were run concurrently with the extraction samples to determine the relative presence of phytoestrogens in the extracts (Fig. 12). Upon viewing the finished plate under UV, however, the standards could not be visualized.



**Figure 12-** Phytoestrogen standards test plate #1 on silica with gypsum. No spots were observed in the standard lane.

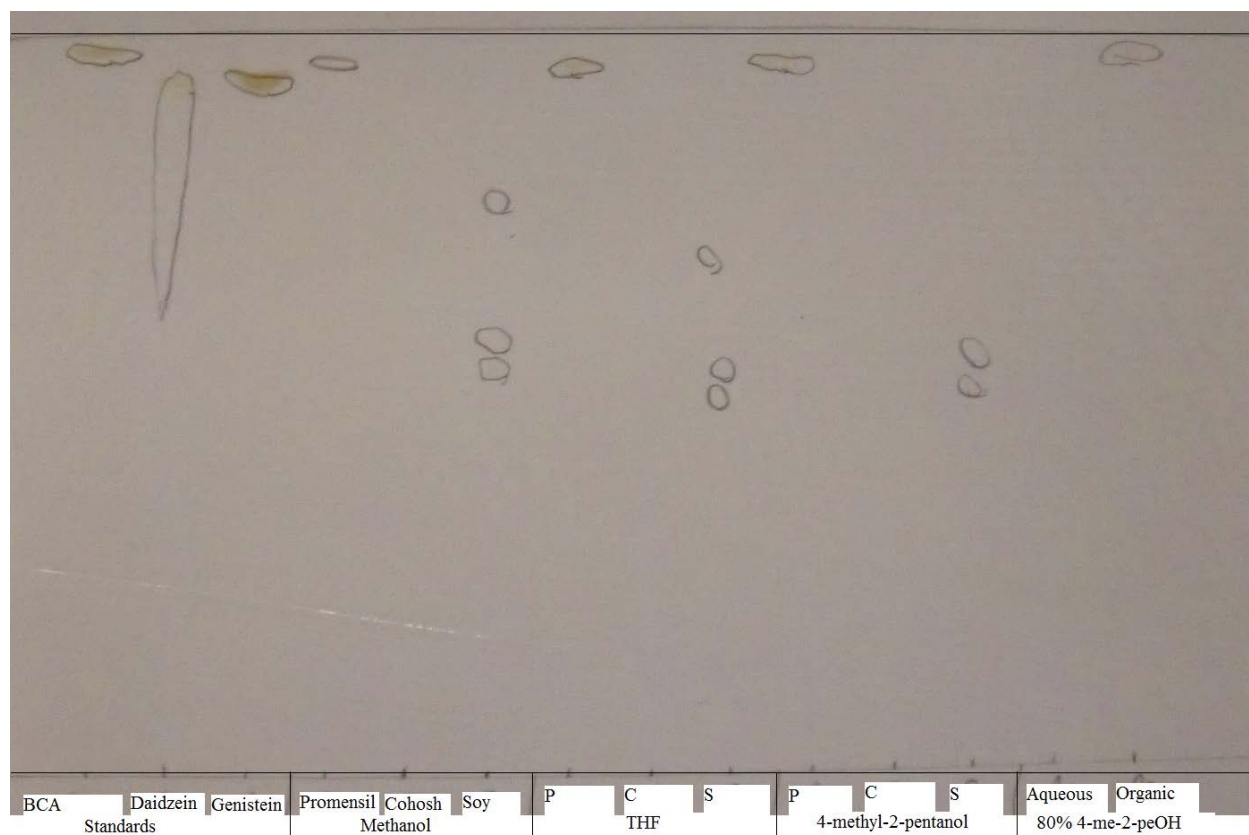
A second test plate was run using UV-active alumina to see if there was any significant difference; this only resulted in the smudging and streaking exhibited in earlier tests using alumina, although there was a smudge showing the presence of daidzein (Fig. 13).



**Figure 13-** Phytoestrogen standards test plate #2 on alumina. Despite excessive smudging of spots, daidzein presence was observed in the phytoestrogen standard lane.

To eliminate further variables, the TLC plates were changed for the same silica F<sub>254</sub> UV-active plates used in Caron's assays (Caron, 2007). One run was carried out to test the new plates (Fig. 14). A small section of one plate was then used for a spot test without being chromatographed to ensure the phytoestrogen standards could be visualized on the plates. All three spots, whether centrifuged or not, could be visualized under short-wave UV (Fig. 15). This confirmed the standards were in fact visible on the silica F<sub>254</sub> plates.





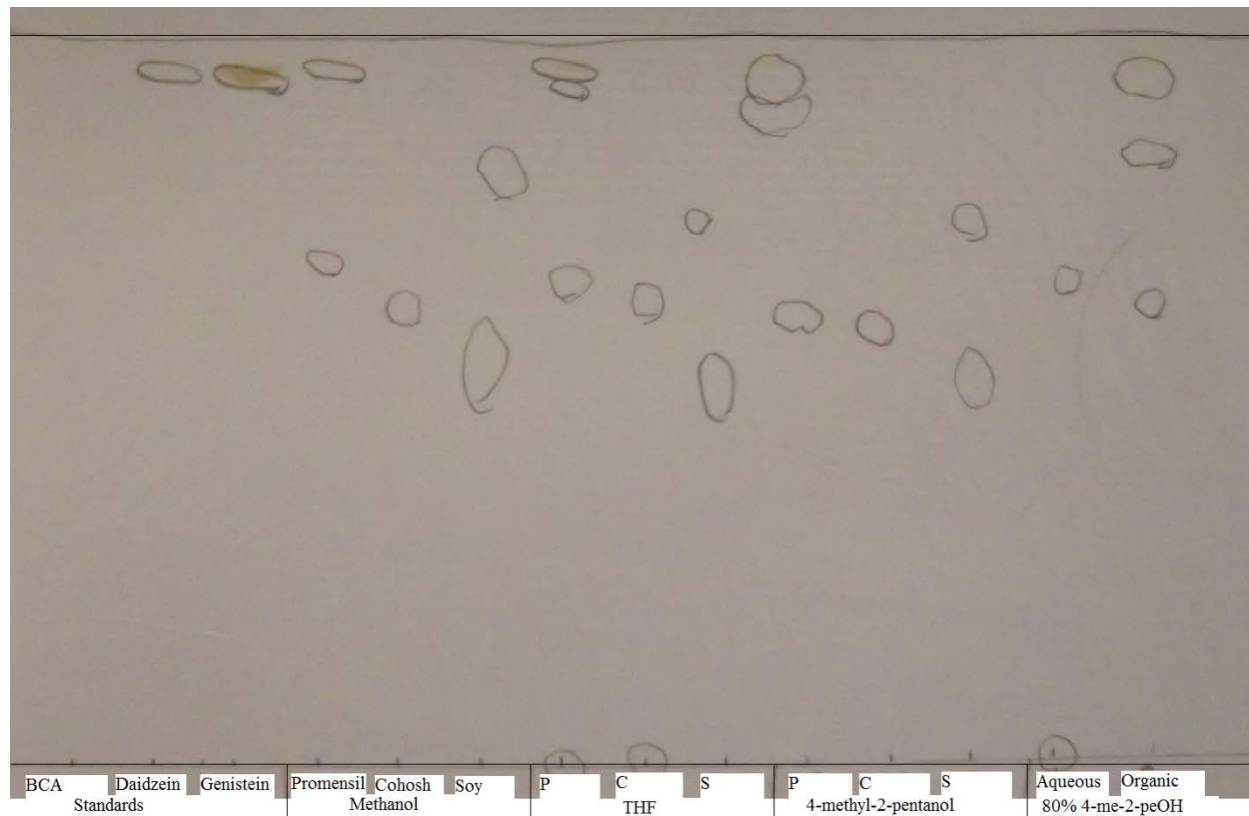
**Figure 14-** Phytoestrogen standards test plate #3 on silica  $F_{254}$ . Spot clarity was significantly improved from that of previous assays.



**Figure 15-** Phytoestrogen standard spot tests. Under short-wave UV, clear spots appeared for all phytoestrogen standards.

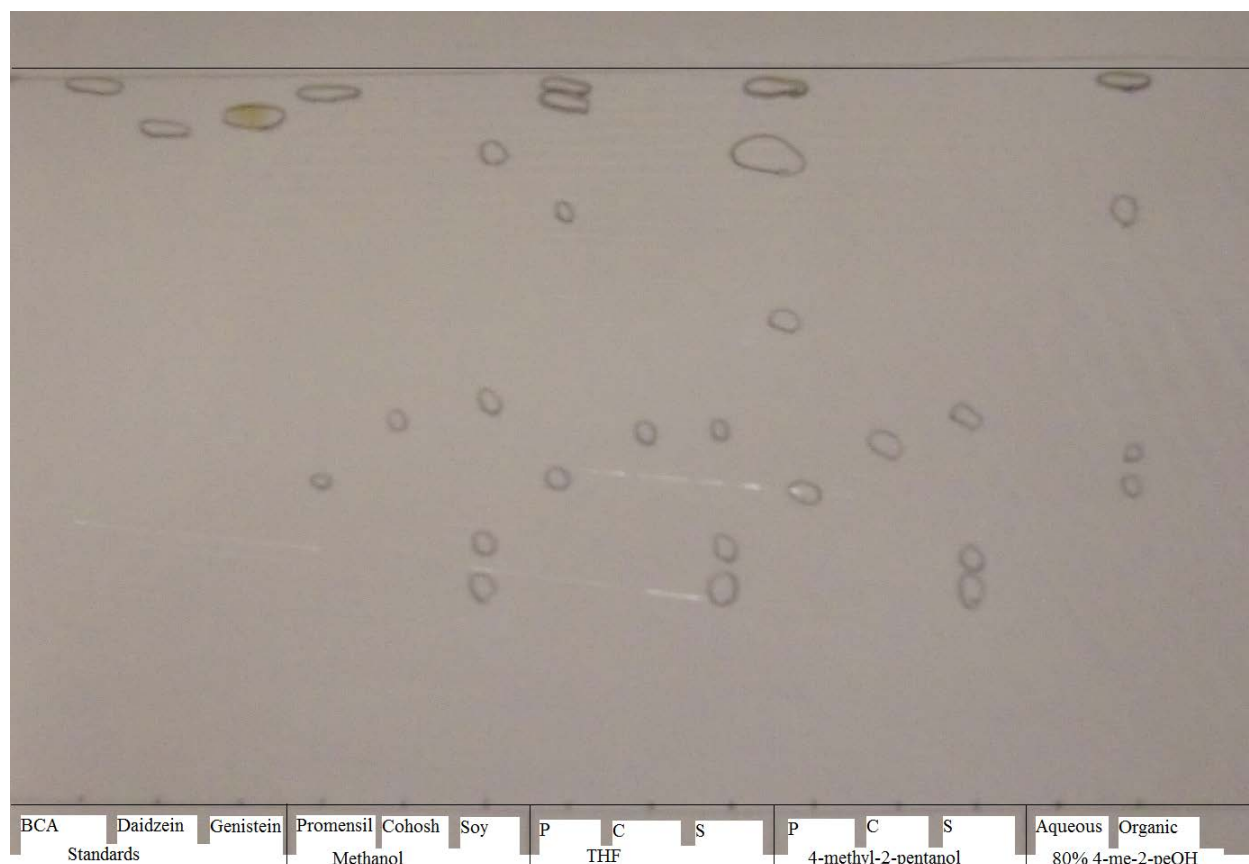


A second test plate was run to confirm the phytoestrogen standards remained visible on the silica F<sub>254</sub> plates after being chromatographed (Fig. 16). These tests determined the standards had been traveling with the solvent front; therefore the solvent system would require modification.



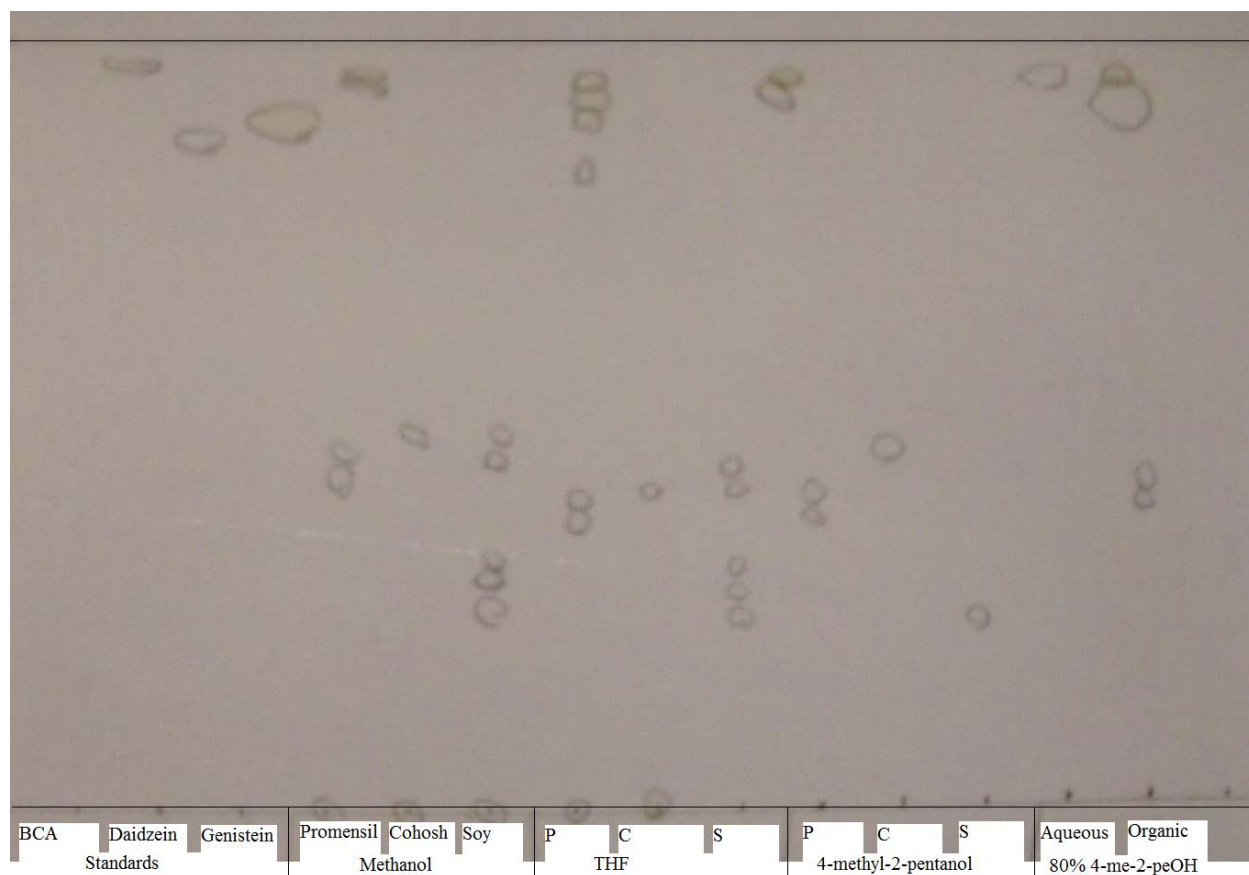
**Figure 16-** Silica F<sub>254</sub>, second test plate. This confirmed the standards were running along with the solvent front and the solvent ratio would require modification.

Following these tests, two full-scale tests were run to modify the relative amount of methanol in the mixture. The first plate was run with an increased ratio of chloroform to create a 12:3:1 chloroform/methanol/acetonitrile mix. Results displayed only a small effect on the movement of the phytoestrogen standards (Fig. 17).



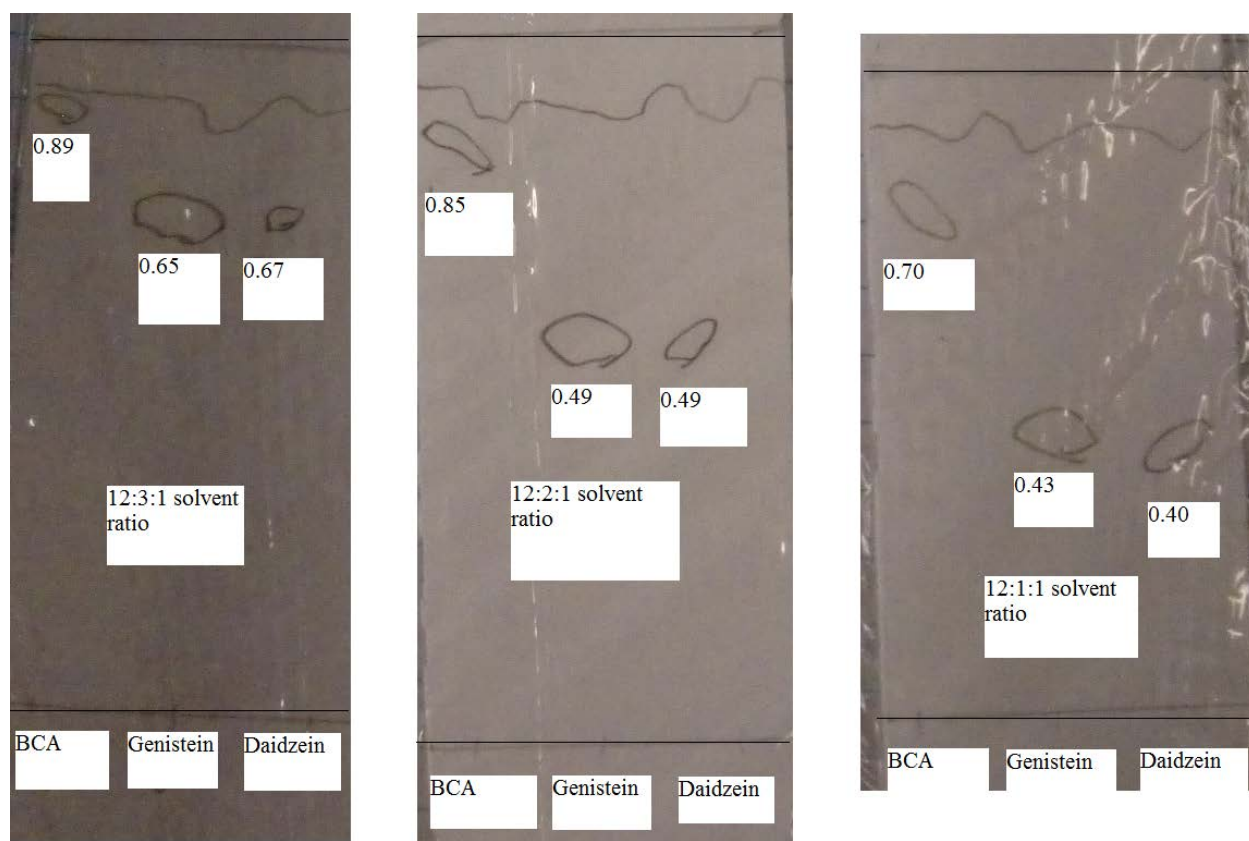
**Figure 17-** Solvent test #1- 12:3:1 chloroform/methanol/acetonitrile. Decreasing the overall ratio of methanol in the solution lowers the interaction of the phytoestrogens with the TLC solvent, effectively slowing their ascent up the plate.

In the second test (Fig. 18), the amount of acetonitrile was increased slightly as well to create a 12:3:2 chloroform/methanol/acetonitrile ratio. The results again displayed a relatively insignificant change, suggesting that a direct decrease of the amount of methanol in the mixture would be more effective than increasing the solvents with lower elutropic values.



**Figure 18-** Solvent test #2- 12:3:2 chloroform/methanol/acetonitrile to further lower the ratio of methanol in the solution. Biochanin A again remained with the solvent front, necessitating more modification of the solvent solution.

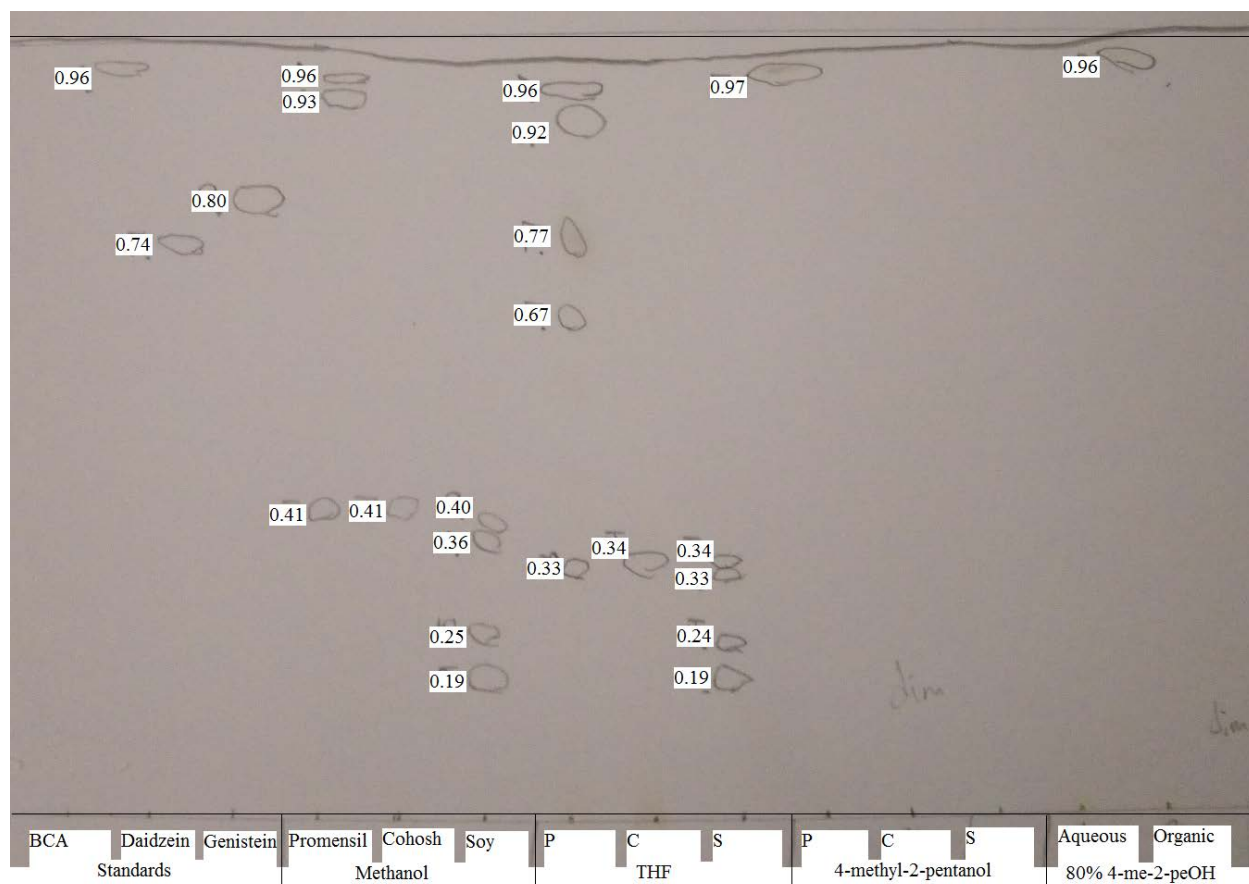
To test the effects of varying amounts of methanol, three small-scale solvent tests were run using small pieces of silica with gypsum plates (Fig. 19). After viewing these results, the 12:2:1 solvent was chosen as the optimal ratio to use in the final full-scale assays.



**Figure 19-** Small-scale solvent tests. From left to right, solvent ratios were: 12:3:1, 12:2:1, and 12:1:1 chloroform/methanol/acetonitrile.

The final TLC method was determined through examination of all previous tests: the 80% methanol, 80% tetrahydrofuran, and pure 4-methyl-2-pentanol extracts would be centrifuged before loading 5  $\mu$ L of each supernatant onto silica F<sub>254</sub> plates. For the 80% 4-methyl-2-pentanol tests, a 5  $\mu$ L sample of both the aqueous and organic phases of the mixture was loaded. Purified standards of biochanin A, genistein, and daidzein were run concurrently. A solvent solution of 12:2:1 chloroform/methanol/acetonitrile was used for the mobile phase, and plates were run for 90 minutes or until the solvent front had traveled more than two-thirds of the way up the plate. Short-wave UV was used to visualize the resultant spots and calculate  $R_f$  values for their identification by comparison with the standards.

The first plate run by this method (Fig. 20) displayed a possible presence of biochanin A in each of the Promensil lanes, with bands appearing around  $R_f = 0.96$  in 4 out of 5 lanes. No other bands appeared consistent with the phytoestrogen standards, although the Promensil extract using tetrahydrofuran solvent exhibited two bands, around  $R_f = 0.77$  and 0.67, which were relatively close to the standard values for genistein at 0.80 and daidzein at 0.74.

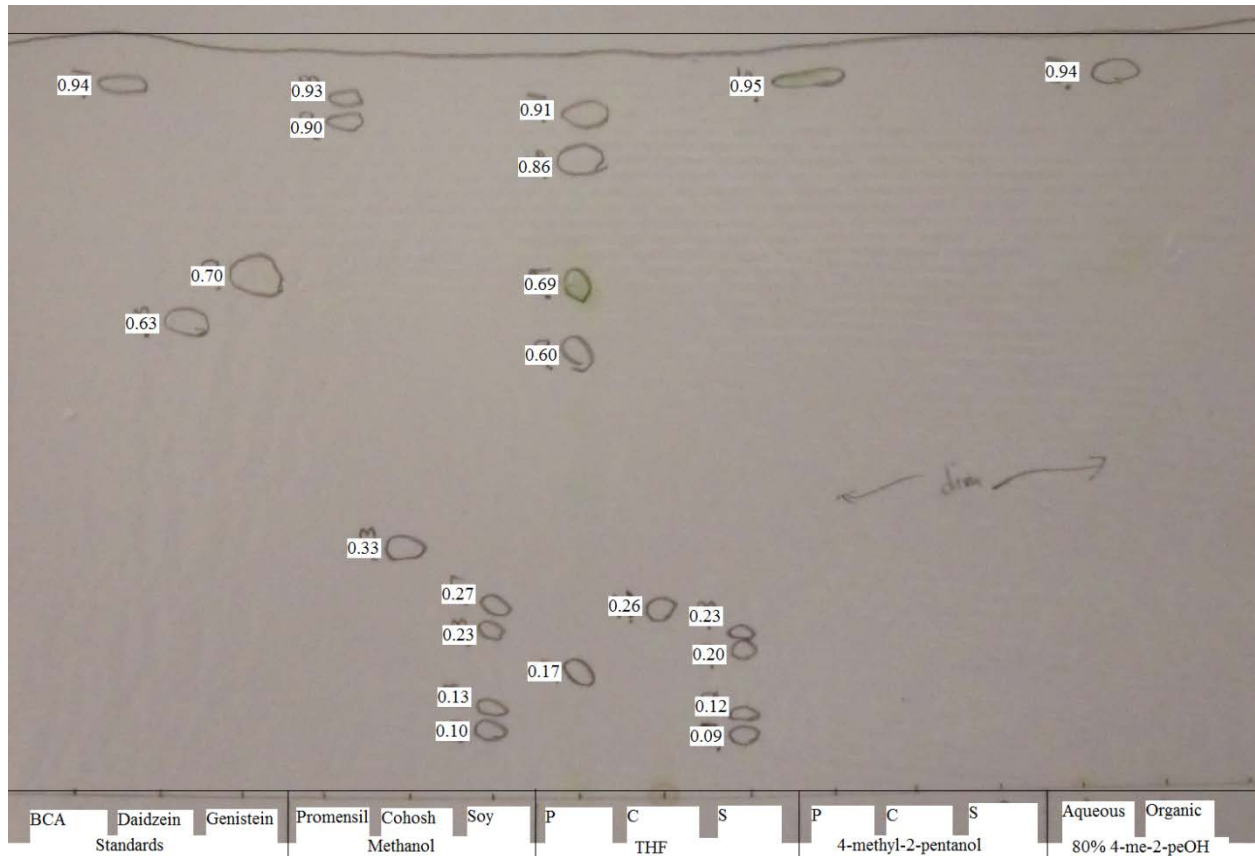


**Figure 20-** Identification of phytoestrogens in extracts, test #1.  $R_f$  values are recorded to the left of their respective bands. The presence of biochanin A is indicated by a spot near 0.96 in each extract.

A second plate was run to check for consistency, and the results were extremely similar (Fig. 21). Most  $R_f$  values were within 0.1 or less from their counterpart bands on the first plate. Again, biochanin A was seen around  $R_f = 0.93$  in each Promensil lane, while the two bands in the THF Promensil lane at 0.69 and 0.60 were even closer to the standard values for genistein at 0.70 and daidzein at 0.63. A compilation of these  $R_f$  values may be seen in Table 1 below.

Lane	Test 1	Test 2	Lane	1	2	Lane	1	2
Biochanin A	0.96	0.94	Daidzein	0.74	0.63	Genistein	0.80	0.70
Methanol Promensil	0.96	0.93	THF Promensil	0.67, 0.77	0.60	THF Promensil	0.77	0.69
THF Pro.	0.96	0.91						
4-methyl-2-pentanol Pro.	0.97	0.95						
80% 4M2P, Aqueous	0.96	0.94						

**Table 1-** Summary of calculated  $R_f$  values corresponding with phytoestrogen standards in Figs. 20 and 21.



**Figure 21-** Confirmation of consistency from phytoestrogen identification test #1, Fig. 20. Presence of biochanin A is observed in each extract; daidzein and genistein seem to be present in the tetrahydrofuran extract of Promensil as well.

These results show tetrahydrofuran may be a more efficient extraction medium than methanol, at least when used for Promensil extractions. However, the bands present which match up to daidzein and genistein standards may only be pigments or other compounds which were extracted by THF and not methanol. Also, past studies using methanol extractions did display small concentrations of genistein present in Promensil extracts (Caron, 2007).

## **Conclusion**

While the extraction of phytoestrogens using methanol has been a useful industry standard for years, it is always prudent to test alternative methods to the norm. Having a large array of techniques at one's disposal allows for more adaptability to experimental conditions as well as unforeseen circumstances. This study demonstrates a possibility for tetrahydrofuran to be used as an extraction solvent for experimentation requiring higher yields of genistein and/or daidzein. Further testing to confirm the presence of these phytoestrogens in the extract, to isolate them from the extract, and to purify them is recommended.

Despite these findings, studies such as those by Setchell *et al* and Caron still indicate methanol to be an overall effective extraction medium for over-the-counter phytoestrogen products (Setchell, 2001; Caron, 2007). When Setchell *et al* analyzed concentrations of phytoestrogens in methanol extracts of Promensil, they did find daidzein and genistein presence as well as biochanin A (Setchell, 2001). Caron's TLC method, which was the basis for this study, also displayed a presence of genistein in the methanol Promensil extract which wasn't observed in this experiment's modified method (Caron, 2007). This may be attributable to the revised method itself or to the high variability of phytoestrogen content in the over-the-counter supplements which was discussed in the introduction.



## CHAPTER II: Separation of Extraction Mixtures by High-Performance Liquid Chromatography

### Introduction

While TLC is a useful method to determine possible phytoestrogen content in over-the-counter supplements, it is best used in conjunction with other analyses such as high-performance liquid chromatography, or HPLC. HPLC provides not only a medium to differentiate between the components of an extraction mixture; it can also be used to analyze the relative concentrations of each component in the extraction. Coward *et al* (1993) were among the earlier groups to analyze phytoestrogens using HPLC, estimating concentrations of daidzein and genistein in high-soy diets such as those typically found in Asian countries. Setchell *et al* (2001) refined the method to be more suitable for analysis of isoflavone supplement extracts and tested 33 over-the-counter products to determine whether or not the supplements contained the amounts of phytoestrogens stated by their manufacturers. It was shown that there were many discrepancies between supplements and the authors recommended more standardization of these products should be enforced (Setchell, 2001). The Setchell *et al* method was later used by Caron (2007) and Flores *et al* (2012) to confirm phytoestrogens' presence in their supplement extracts before testing them on live cell lines.

It was briefly mentioned in the first chapter that anti-proliferative effects on cancer cells have been observed upon their exposure to certain phytoestrogens and isoflavone supplements. Caron's thesis (2007) shows a marked decrease in growth of MCF7 breast cancer cell line upon exposure to extracts from the over-the-counter supplement Promensil. Later, purified standards of phytoestrogens were tested on a T47D breast cancer cell line and it was observed that resveratrol and biochanin A were particularly effective at inducing cell apoptosis (Comeau, 2010). Further experimentation by Park and Patchel (2011) and Flores *et al* (2012) tested two other supplements, one derived from black cohosh and one from soy plants, for their effects on cell proliferation. Again Promensil displayed significant anti-proliferative qualities when tested on the MCF7 cell line; when exposed to the extract, cell counts decreased by 30-55% when compared to controls treated with only methanol. The black cohosh extract was observed to be effective as well, reducing cell growth by 35-60%. Soy extract was the weakest of the three: between the two studies it lowered cell counts anywhere from 0-35% of the control. Similar results for black cohosh and soy were obtained upon testing T47D cell lines, but oddly enough in preliminary results Promensil seemed to have little effect (Flores, 2012).

The studies by Park and Patchel (2011) and Flores *et al* (2012) lead one to wonder what may be the cause of the differential effects observed in their three isoflavone supplements; is it simply a matter of differing phytoestrogen concentrations or are certain phytoestrogens more effective in their anti-proliferative qualities than others? Perhaps other compounds may be



present in Promensil which amplify the anti-proliferative effects of its phytoestrogens, or it may lack certain compounds in soy and black cohosh which inhibit these effects. Whatever the case, the first step to answering these questions lies in determining the differences between the three extraction mixtures through HPLC.

## **Materials and Methods**

### **Analysis by HPLC:**

HPLC was carried out on a Hewlett-Packard 1100 apparatus using the method described in Setchell et al (2001) and Caron (2007). Using a PerSeptive Biosystems reverse phase PepMap C18 column, 10  $\mu$ L of sample or standard were injected at a flow rate of 1.0 mL/min. The column was then re-equilibrated in 10 mM ammonium acetate (0.1% trifluoroacetic acid [TFA]) for 2 minutes. Elution followed for 22 minutes using a linear gradient of 10 mM ammonium acetate (0.1% TFA) 100-50% and acetonitrile 0-50% until the column was held isocratic at 50/50 for 5 minutes. The column was then re-equilibrated using 10 mM ammonium acetate (0.1% TFA). Absorbance was measured at 260 nm for the entire process. 100  $\mu$ M pure standards of genistein, daidzein, and biochanin A were prepared in methanol and run separately to determine their respective retention times. The extraction mixtures were centrifuged at 3000 RPM for 5 minutes and the supernatant passed through a 0.8  $\mu$ m syringe filter to eliminate particulates before being run.

As detailed in Chapter I, supplement extractions were carried out by refluxing each of Promensil, soy, and black cohosh supplements in 80% methanol for one hour. The resulting mixtures were filtered and used as the three HPLC extract samples.

Hewlett-Packard ChemStation software for liquid chromatography systems was used for data analysis and determining integration values of HPLC traces.

## **Results and Discussion**

*From this point on, HPLC data will be presented and discussed using only specific traces selected from three runs per each of three phytoestrogen standards and three extract samples. To see the full set of traces as well as integration data for each, please refer to the Appendix.*

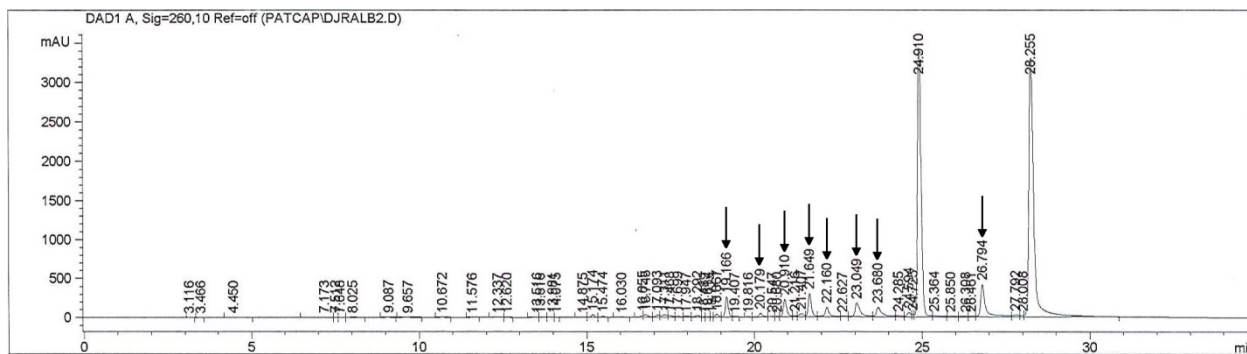
The three pure phytoestrogen standards were each run in triplicate to confirm consistency of the observed peak and calculate average retention times for each standard. These results may be seen in Table 2 below.

<b>Standard</b>	<b>Run 1 RT (min.)</b>	<b>Run 2 RT (min.)</b>	<b>Run 3 RT (min.)</b>	<b>% Variance</b>	<b>Avg. RT (min.)</b>
Daidzein	20.061	20.120	20.106	0.29	20.096
Genistein	22.956	23.060	22.965	0.45	22.994
Biochanin A	28.279	28.310	28.220	0.32	28.270

***Table 2-**Retention times of three phytoestrogen standards. The standards were prepared to 100  $\mu$ M in methanol. Percent variance between the highest and lowest RT for each standard was calculated to confirm consistency.*

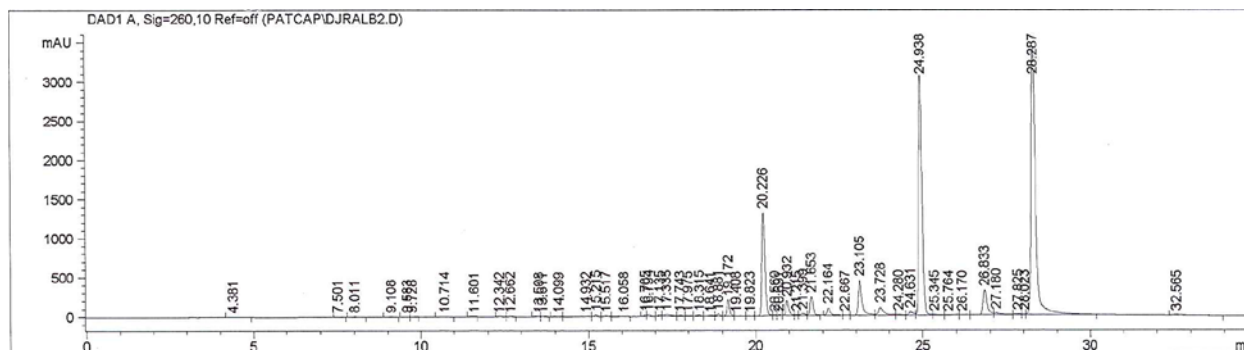
All standards had a variance of less than 0.5% among three runs, confirming precision of the method and apparatus used. For later comparison to the phytoestrogen extracts average retention times were calculated to be 20.096 min. for daidzein, 22.994 for genistein, and 28.270 for biochanin A. Expected retention times for daidzein, genistein, and biochanin A were derived from Setchell (2001), Caron (2007), and Flores (2012). These were estimated at 21, 23, and 29 minutes respectively. Thus the observed average retention times were earlier than expected, but consistently so; this can be attributed to the use of a different apparatus or HPLC column than the previous experiments. Due to the low calculated percent error the difference is negligible.

Upon examination of the Promensil extract traces, the peaks with retention times most comparable to the standards were found at 20.179, 23.049, and 28.255 minutes respectively. It is assumed these peaks represent daidzein, genistein, and biochanin A presence. The biochanin A peak was especially pronounced. Taking the results of Comeau and Skorinko into consideration, it is likely that biochanin A plays a large part in the observed effectiveness of Promensil in reducing cell proliferation (Comeau, 2010). An unknown major peak was also observed at 24.910 minutes. Other peaks of interest, whose percent areas after integration were greater than one, occurred at retention times of 19.166, 20.910, 21.649, 22.160, 23.680, and 26.794 minutes. These results may be seen in Figure 22 below.



**Figure 22** – HPLC trace of Promensil extract. Major peaks are seen at retention times of 24.910 and 28.255 minutes. Arrows indicate peaks of interest with areas greater than one percent.

To confirm the identity of the three possible phytoestrogen peaks, a second Promensil sample was spiked with the pure phytoestrogen standards and run through HPLC. This result may be seen below (Fig. 23).



**Figure 23** – HPLC trace of Promensil extract spiked with pure standards of daidzein, genistein, and biochanin A. The three peaks of interest from the base Promensil extract were analyzed, and two of three displayed an increased % area over the initial Promensil runs. The peak at 28.287, corresponding with biochanin A, was off the scale and thus no difference could be observed.

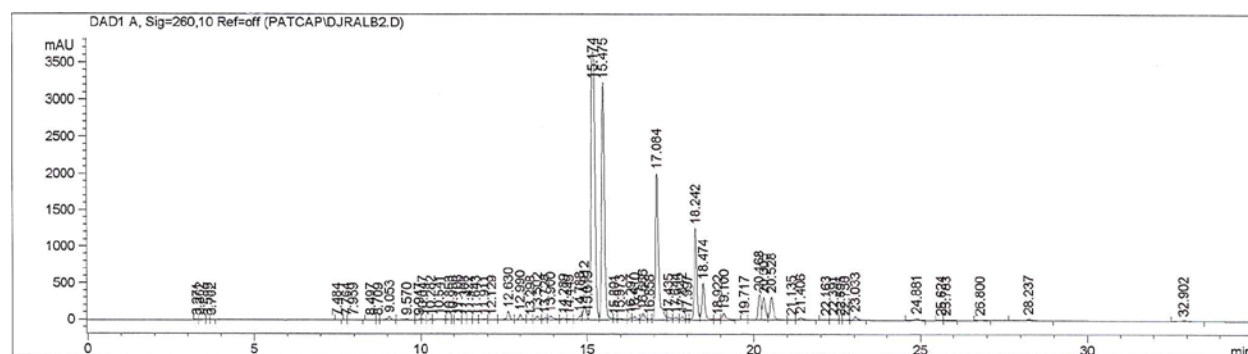
Average retention times and percent areas of the relevant peaks in both the initial Promensil extract runs and the spiked runs were calculated for comparison between the two. Results displayed a ratio greater than one between percent areas of the spiked and unspiked mixtures for both peaks representing daidzein and genistein near 20.096 and 22.994 respectively; this data as well as there being no change in the peaks' shape confirms the identity and presence of daidzein and genistein in the Promensil extract. The peak at 28.287 of the spiked mixture exhibited a similar percent area to the corresponding peak on the initial Promensil run rather than increasing, but this discrepancy may be ignored since both signals were too strong for the detector. There was no change in the shape of this peak so it will be assumed it represents a high concentration of biochanin A. This corresponds with the findings of Setchell *et al*: their test of Promensil displayed a high concentration of biochanin A in the extract as well (Setchell, 2001). For further confirmation one could dilute the Promensil extract to weaken the signal at 28.287 then spike the diluted mixture with pure biochanin A standard, but this was deemed unnecessary for this study.

after considering the results of Setchell *et al.* Data from comparison of the initial Promensil extract and the spiked extract may be seen in Table 3 below.

Compound	Avg. Standard RT (min.)	Nearest Peak RT (min.)	Peak % Area	Spiked Nearest Peak RT (min.)	Spiked Peak % Area	Area Ratio (spiked/ unspiked)
Daidzein	20.096	20.171	0.4839	20.226	9.9903	20.65
Genistein	22.994	23.039	2.4614	23.108	4.7160	1.916
Biochanin A	28.270	28.243	41.928	28.289	40.948	0.9766

**Table 3** – Analysis of HPLC data between initial Promensil extract runs and Promensil spiked with phytoestrogen standards. Ratios increased when the spiked peak percent areas of daidzein and genistein peaks were compared to the corresponding initial Promensil extract peak percent areas, confirming the identity of those peaks.

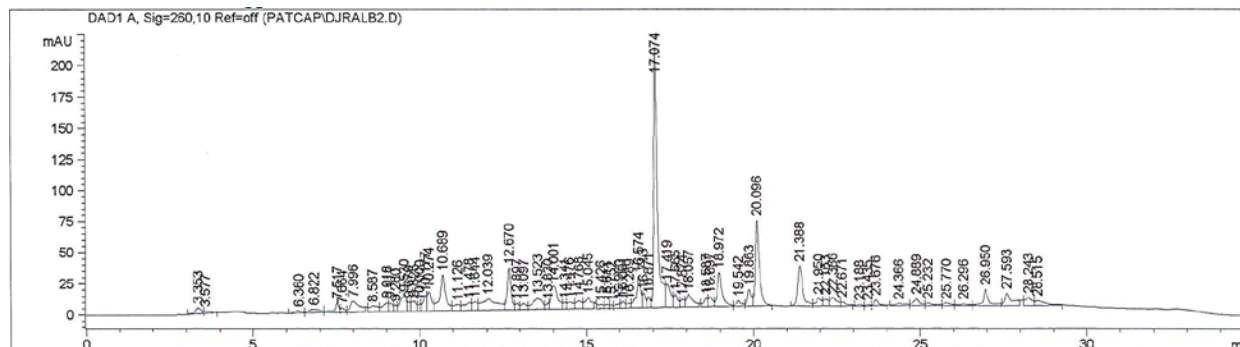
The soy extract (Fig. 24) exhibited exceedingly small peaks at the three phytoestrogen standard retention times. One peak at 20.168 minutes may coincide with daidzein; genistein and biochanin A peak areas were so small as to be negligible when compared to other components of the mixture. Larger, unidentified peaks were observed at 15.174, 15.475, 17.084, 18.242, and 18.474. Other notable peaks occurred at 14.912, 20.305, and 20.528 minutes.



**Figure 24** – HPLC trace of soy extract. Several major peaks were observed; none coincided with phytoestrogen standard RTs.

Black cohosh extract (Fig. 25) yielded a relatively large peak at 20.096 minutes, coinciding exactly with the calculated retention time for daidzein. A small peak at 28.243 displays possible biochanin A content as well; as with Promensil, this may contribute to the black cohosh extract's observed anti-proliferative effects in previous tests (Park, 2011; Flores, 2012). One small peak at 23.188 suggests low genistein concentrations. Major peaks were also observed at 17.074 and 21.388 minutes. Other peaks of interest occurred at 7.996, 8.587, 9.018, 9.530, 10.177, 10.274, 10.689, 11.478, 12.039, 12.670, 13.523, 14.001, 15.045, 16.574, 17.419, 17.665, 18.057, 18.972, 19.863, 22.386, 26.950, 27.593, and 28.515 minutes. The high incidence of this many peaks displaying areas greater than one percent suggests the anti-

proliferative effects of the black cohosh mixture may be attributable not only to its phytoestrogen content, but also to an efficient combination of many constituents. Results may be seen below.



**Figure 25** –HPLC trace of black cohosh extract. The major peak at 20.096 coincides exactly with the calculated average retention time for daidzein.

In a cursory visual comparison of the three extracts it quickly becomes apparent there are significant differences in retention times between their major peaks. One's eye is drawn to the peaks occurring at 24.910 and 28.255 for Promensil; at 17.074, 20.096, and 21.388 for black cohosh; and at 15.174, 15.475, 17.084, 18.242, and 18.474 for soy. Even without referencing the integration data the height of these peaks causes them to stand out among the traces. Closer examination of integration data involving all peaks of interest shows several unique peaks in each extract. The major peak at 24.910 of the Promensil extract stands out in particular with a strong reading that exceeds the scale of the trace, but peaks of interest at 19.166, 20.910, 23.680, and 26.794 are also unique to Promensil.

The soy extract was different in that all its peaks of interest occurred before 21 minutes, effectively ruling out genistein at 22.944 and biochanin A at 28.270. The presence of daidzein was likely with a peak observed at 20.168. One of soy's major peaks at retention time 17.084 coincided with a strong black cohosh peak at 17.074. Considering the large disparity between the anti-proliferative effects of soy and black cohosh (Park, 2011; Flores, 2012), it is possible to rule out these peaks as having little to no influence on the anti-proliferative effects of the extracts as a whole. The remaining peaks of interest unique to soy occur at 14.912, 15.174, 15.475, 18.242, 18.474, 20.305, and 20.528 minutes.

Black cohosh displayed more than twice as many peaks with areas greater than one percent when compared to the other two extracts. The three strongest signals were measured at 17.074, 20.096, and 21.388. As mentioned in the previous paragraph, the peak at 17.074 coincides with a large peak in soy and is therefore ruled out of the equation when considering the differential effects between these mixtures. The peak at 20.096 coincides precisely with the calculated retention time for the pure daidzein standard. Of particular interest is the peak

occurring at 21.388, which is unique to the black cohosh extract. Since the peaks of interest were so numerous in the black cohosh extract, there are myriad others which were unique to the mixture; those occurring at 10.689, 12.670, and 18.972 demonstrated the strongest signals of the remaining peaks.

The assumed common presence of biochanin A in both Promensil and black cohosh suggests the compound may have the strongest anti-proliferative effect of the three phytoestrogen standards tested since both extracts exhibited such strong effects on live cell lines (Park, 2011; Flores, 2012). The soy extract exhibited a negligible peak at the corresponding retention time, possibly accounting for it having the weakest effect from among the three supplements.

Possible presence of daidzein was observed in varying concentrations in all three extracts. This indicates a possibility that any anti-proliferative effects of daidzein may be highly dependent on concentration, as the lowest concentration occurred in the relatively ineffective soy product. Another possibility is that daidzein works more efficiently in tandem with biochanin A, which is absent from the soy extract.

## **Conclusion**

The intent of this second chapter was to determine unique components between the three phytoestrogen supplement extracts which would help to explain the differences in their effects on breast cancer cell proliferation. While there were several constituents unique to each mixture, those present in the soy extract were somewhat less intriguing due to its weak observed anti-proliferative effects (Park, 2011; Flores, 2012). That being said, unless more positive results are found for the soy extract in the future it may not be a worthwhile pursuit for anti-proliferation testing. Phytoestrogen content in the soy was also very low among the three standards run for comparison, although this may simply be a result of the product being unregulated. The results of Coward *et al* reinforce this fact; natural intake of soy in one's diet would seem to be more effective than this soy supplement in certain instances (Coward, 1993). Perhaps there is more to the soy diet's success than its phytoestrogen content.

The significant anti-proliferative effects of the Promensil and black cohosh extracts may be attributable to their combinations of phytoestrogens, but both extracts exhibited their own strong unique peaks as well which may be contributing factors in their effectiveness. Promensil seems to contain a larger concentration of biochanin A, while black cohosh displayed more daidzein. Tests on breast cancer cell lines involving combinations of daidzein and biochanin A in various ratios would be helpful to determine whether or not these compounds have a greater anti-proliferative effect in tandem than they do alone. Isolation and analysis of the major Promensil peaks at 24.910 and 28.255 is strongly recommended for further testing on live cell lines, as well as the major black cohosh peak at 21.388.

Setchell *et al* found biochanin A to be the largest phytoestrogen component of Promensil by far (44,330 µg/g), which helps explain why the signal strength in this study's extract was far too strong for the HPLC spectrometer to measure correctly. Also occurring in a high concentration was the isoflavone formononetin at a concentration of 26,726 µg/g; this may account for the large unknown peak on the Promensil trace at 24.910. Daidzein content was calculated at 1,532 µg/g and genistein at 2,900 µg/g (Setchell, 2001). Here the variable nature of these over-the-counter supplements may be seen again; the HPLC signals observed from this study's Promensil extract indicated a slightly higher concentration of daidzein rather than genistein in the extract.

Another point to take into consideration when discussing the anti-proliferative effects of phytoestrogens is their possible metabolism when introduced into the human body. Due to their structural similarities to human estrogens it's entirely possible phytoestrogens may be broken down into metabolite forms as well. This may explain the differential effects observed in natural soy diets over soy isoflavone supplements due to the difference in methods of delivery (Coward, 1993). While the compounds themselves seem to have effects on cancer cell proliferation when



introduced directly to cell lines via petri dishes, it would seem to be more difficult for them to stay intact and reach their desired destination in the body when they must be ingested and first travel through the stomach to the bloodstream to be distributed. With the high likelihood of some sort of metabolism taking place along the way, phytoestrogens from supplements would not arrive at the breast epithelium in a pure form but instead in a mixture of metabolites which may induce different effects. Future studies could be developed on this basis; perhaps phytoestrogen standards could be treated with estrogen-metabolizing enzymes and then added to cancer cell lines to observe any differential effects from the pure compounds.

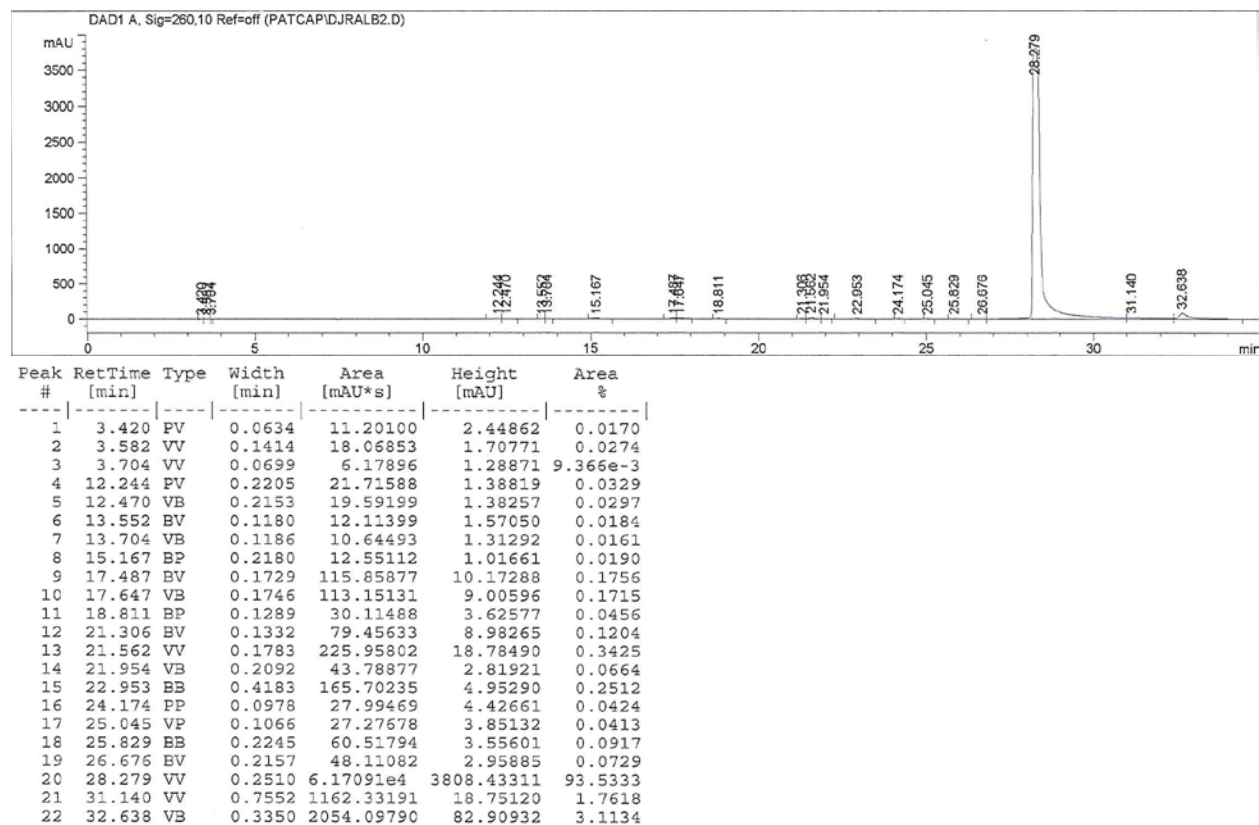
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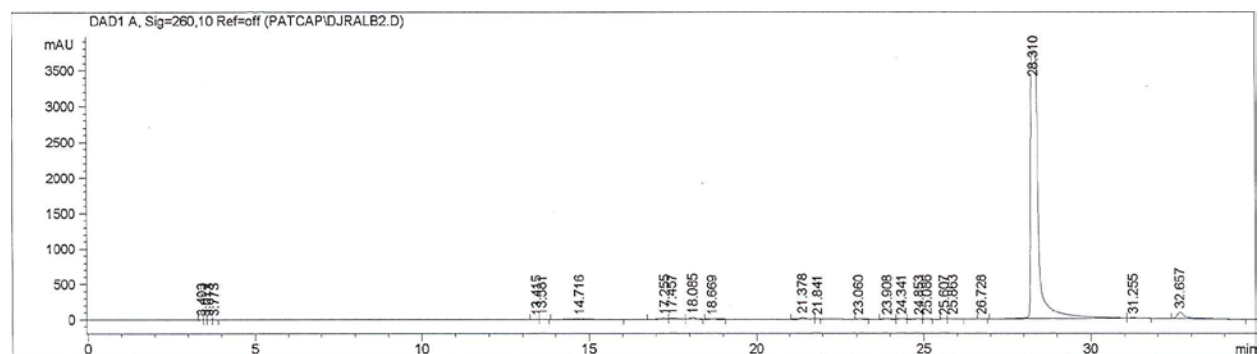
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## Appendix of HPLC Data

### Biochanin A: 1

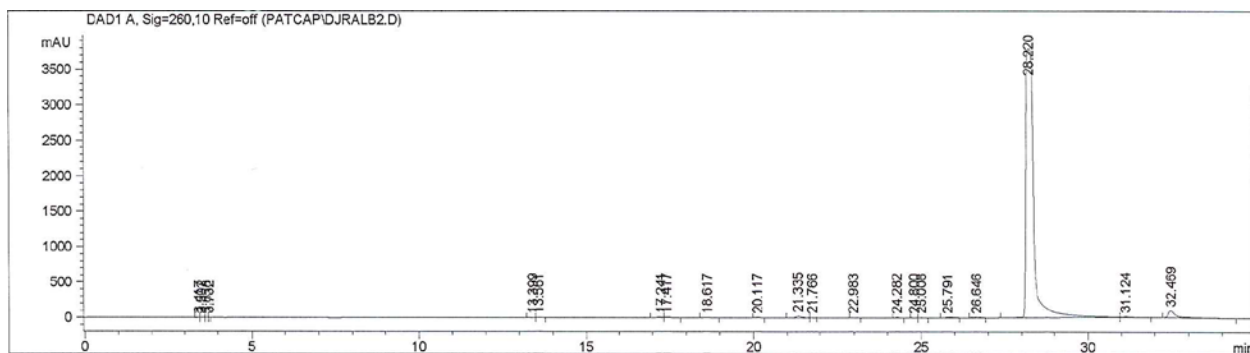


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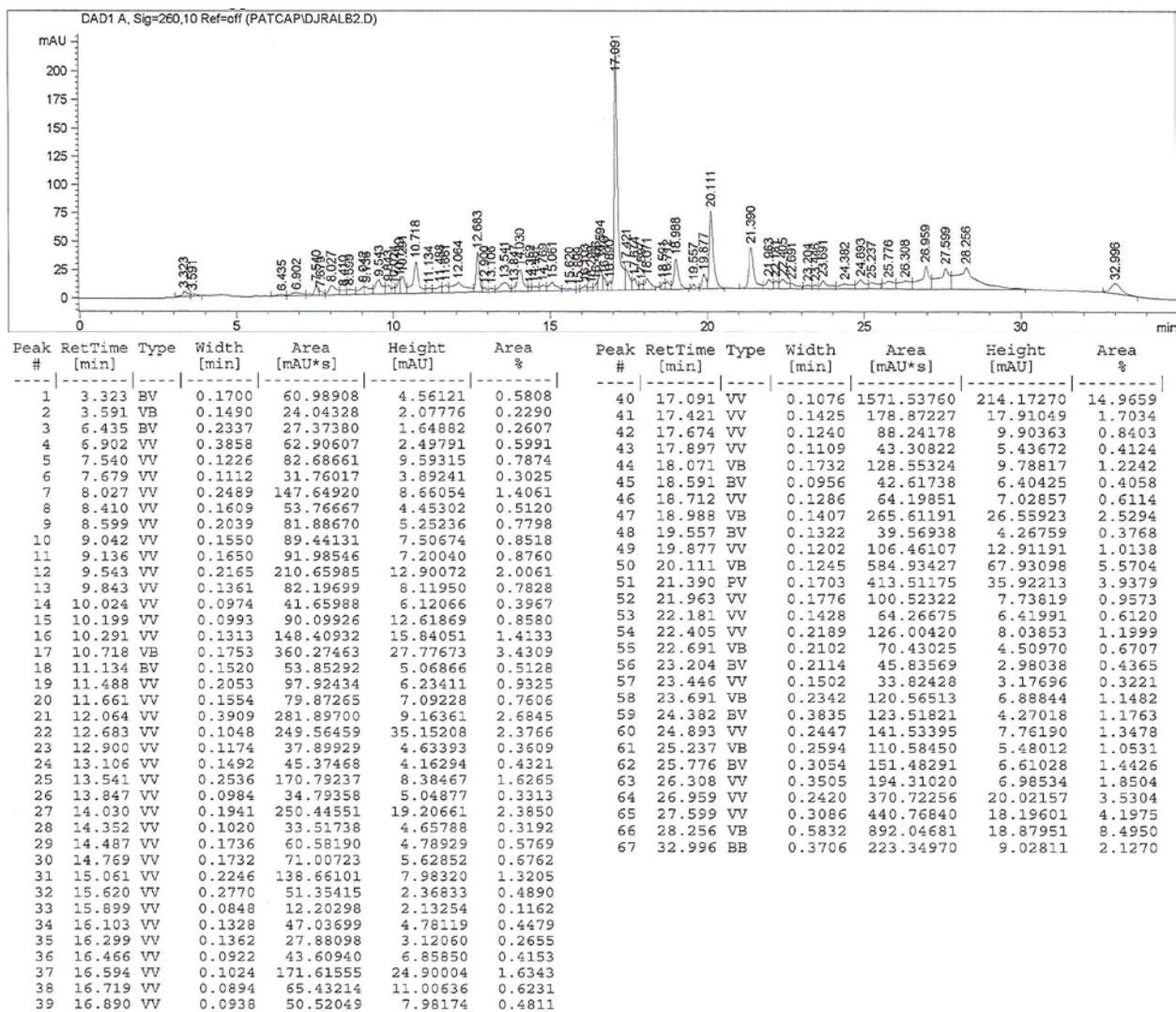
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.403	PV	0.0880	18.07241	3.01090	0.0278
2	3.517	VV	0.0843	14.98076	2.41641	0.0230
3	3.613	VV	0.1021	14.04636	1.85992	0.0216
4	3.773	VB	0.0987	9.01611	1.30325	0.0139
5	13.415	BV	0.1388	16.45853	1.79722	0.0253
6	13.581	VB	0.1603	18.77824	1.58418	0.0289
7	14.716	BB	0.6954	351.55414	6.41434	0.5402
8	17.255	BV	0.2597	223.31731	12.78981	0.3432
9	17.457	VV	0.2783	267.37350	12.77113	0.4109
10	18.085	VB	0.1696	315.82928	27.58400	0.4853
11	18.669	BB	0.2409	48.16734	2.97457	0.0740
12	21.378	PB	0.1943	298.60202	24.47640	0.4588
13	21.841	BV	0.1131	8.24754	1.10448	0.0127
14	23.060	BB	0.1310	17.67395	1.92648	0.0272
15	23.908	BP	0.2630	16.79512	1.01528	0.0258
16	24.341	VB	0.1013	36.13482	5.45036	0.0555
17	24.853	BV	0.1708	18.50546	1.47080	0.0284
18	25.086	VP	0.0990	23.28753	3.62461	0.0358
19	25.607	PV	0.0962	9.64060	1.55880	0.0148
20	25.863	VP	0.1749	45.92624	3.85858	0.0706
21	26.728	BB	0.1450	19.95387	1.92359	0.0307
22	28.310	BV	0.2516	6.13781e4	3776.39771	94.3159
23	31.255	VB	0.2738	208.63383	10.41256	0.3206
24	32.657	BP	0.2628	1698.05786	90.52945	2.6093

### Biochanin A: 3

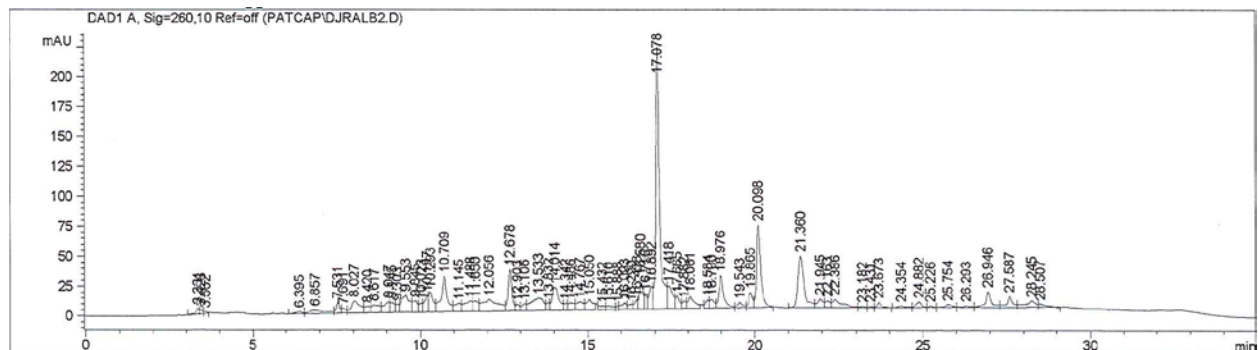


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.417	PV	0.0888	20.39213	3.35997	0.0320
2	3.502	VV	0.0995	20.93274	2.79210	0.0329
3	3.658	VV	0.0744	10.52995	2.03288	0.0165
4	3.732	VV	0.0539	5.68595	1.51135	8.929e-3
5	13.399	BV	0.1334	13.01288	1.49660	0.0204
6	13.561	VB	0.1275	11.77038	1.32616	0.0185
7	17.241	BV	0.2005	109.33137	8.58887	0.1717
8	17.417	VB	0.1918	102.68456	7.49797	0.1612
9	18.617	PB	0.2025	31.64322	2.48549	0.0497
10	20.117	BB	0.1193	50.81451	6.49377	0.0798
11	21.335	PB	0.1993	297.13226	23.53485	0.4666
12	21.766	BV	0.1237	12.76559	1.55572	0.0200
13	22.983	PB	0.1204	16.37828	2.02504	0.0257
14	24.282	PB	0.0997	33.27325	5.13012	0.0523
15	24.800	PV	0.1025	7.95121	1.18216	0.0125
16	25.008	VP	0.1049	24.83398	3.67128	0.0390
17	25.791	BB	0.1871	47.93239	3.80219	0.0753
18	26.646	PB	0.1527	22.39539	2.06245	0.0352
19	28.220	BV	0.2493	6.08508e4	3790.10913	95.5566
20	31.124	VB	0.3156	287.58640	12.17243	0.4516
21	32.469	BP	0.2473	1702.52307	97.69281	2.6735

## Black Cohosh: 1



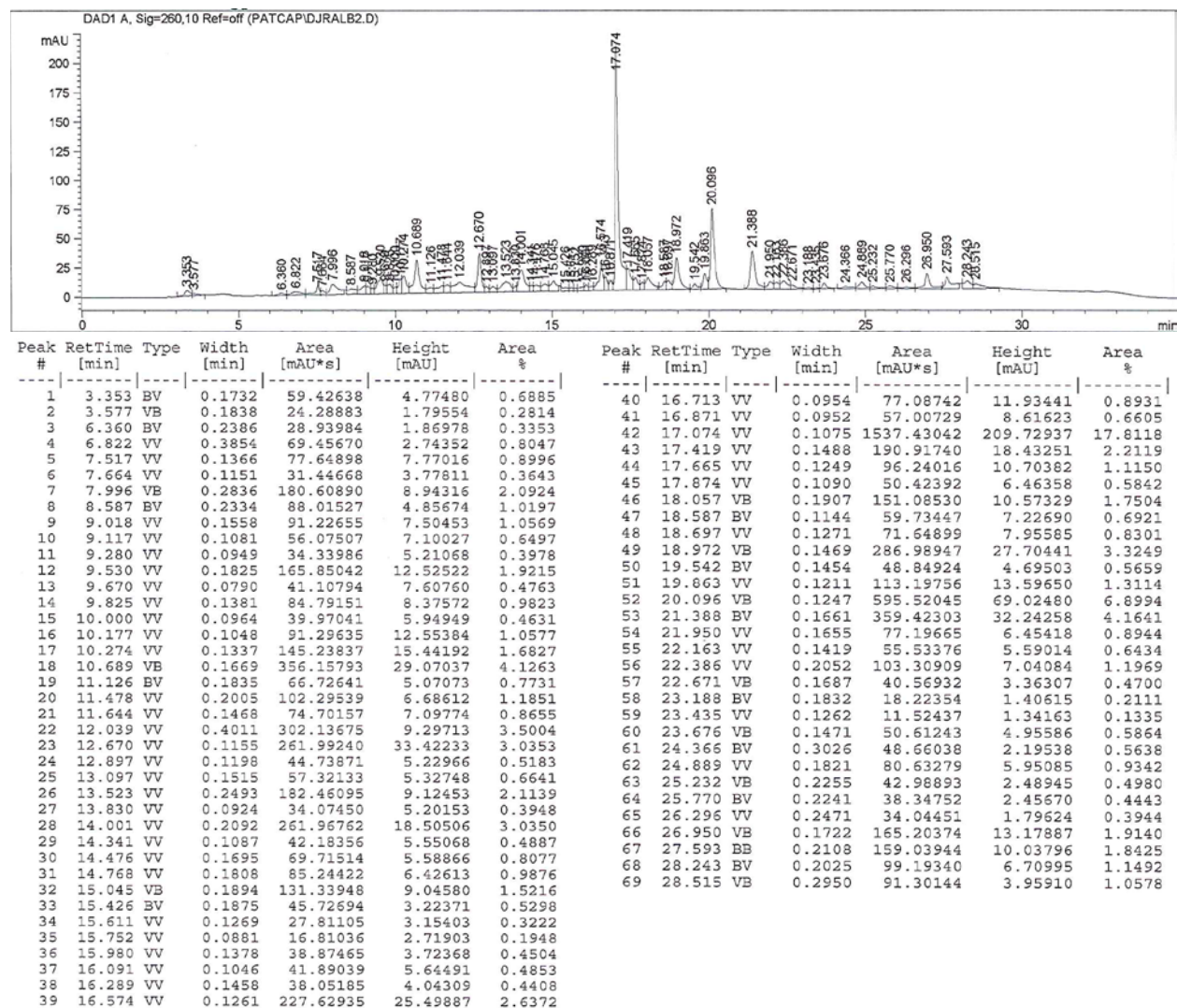
## Black Cohosh: 2



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %	Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.331	BV	0.1237	39.30570	4.69513	0.4122	40	16.580	VV	0.1041	182.98045	26.01213	1.9187
2	3.428	VV	0.0791	20.64158	3.69832	0.2164	41	16.716	VV	0.0909	81.44489	13.02529	0.8540
3	3.562	VV	0.0948	12.42051	1.75054	0.1302	42	16.892	VV	0.1088	143.91362	19.35254	1.5091
4	6.395	BV	0.2464	28.55786	1.75012	0.2995	43	17.078	VV	0.1122	1695.89709	219.34052	17.7833
5	6.857	VB	0.4750	94.78476	2.82786	0.9939	44	17.418	VV	0.1510	207.11124	19.33635	2.1718
6	7.531	BV	0.1076	64.50724	9.00755	0.6764	45	17.665	VV	0.1220	96.19509	11.00613	1.0087
7	7.691	VV	0.1226	36.76585	4.10122	0.3855	46	17.882	VV	0.1110	51.56202	6.46776	0.5407
8	8.027	VV	0.2657	185.98834	10.05992	1.9503	47	18.061	VB	0.1868	149.84564	10.61003	1.5713
9	8.420	VV	0.1552	62.71326	5.41073	0.6576	48	18.584	BV	0.1018	53.30073	7.41695	0.5589
10	8.617	VV	0.2177	107.87637	6.43199	1.1312	49	18.700	VV	0.1303	73.94314	7.96646	0.7754
11	9.047	VV	0.1672	117.29130	9.05272	1.2299	50	18.976	VB	0.1447	286.19873	27.65849	3.0011
12	9.145	VV	0.1103	70.78458	8.76157	0.7423	51	19.543	BV	0.1460	49.26227	4.70937	0.5166
13	9.303	VV	0.0937	46.35076	6.95603	0.4860	52	19.865	VV	0.1211	111.98000	13.45127	1.1742
14	9.553	VV	0.2260	248.41360	14.48875	2.6049	53	20.098	VB	0.1242	592.30304	68.93871	6.2109
15	9.845	VV	0.1401	102.97150	10.00381	1.0798	54	21.360	PB	0.1925	551.45453	43.31725	5.7826
16	10.023	VV	0.0972	51.70028	7.61819	0.5421	55	21.945	BV	0.1718	95.34554	7.62968	0.9998
17	10.197	VV	0.1055	106.50985	14.20583	1.1169	56	22.163	VV	0.1396	55.57648	5.70976	0.5828
18	10.293	VV	0.1377	167.62688	17.19443	1.7577	57	22.386	VB	0.2725	144.10449	7.04849	1.5111
19	10.709	VV	0.1780	393.66776	30.20677	4.1280	58	23.182	BV	0.1690	15.03723	1.29949	0.1577
20	11.145	VV	0.1910	97.77959	7.08767	1.0253	59	23.431	VV	0.1145	8.85270	1.16674	0.0928
21	11.488	VV	0.2206	151.97136	8.92864	1.5936	60	23.673	VB	0.1298	40.70549	4.66391	0.4268
22	11.650	VV	0.1484	95.85345	8.84721	1.0051	61	24.354	FV	0.2456	27.16561	1.63746	0.2849
23	12.056	VV	0.4088	333.12244	10.15201	3.4932	62	24.882	VV	0.1617	56.06490	4.82029	0.5879
24	12.678	VV	0.1130	266.59232	34.94928	2.7955	63	25.226	VV	0.1275	14.73017	1.65978	0.1545
25	12.903	VV	0.1217	48.63943	5.69382	0.5100	64	25.754	VV	0.2279	42.54084	2.64097	0.4461
26	13.106	VV	0.1249	54.14741	6.02239	0.5678	65	26.293	VV	0.2122	26.62823	1.66824	0.2792
27	13.533	VV	0.3132	283.22906	11.11851	2.9700	66	26.946	VV	0.1674	164.07671	13.54009	1.7205
28	13.832	VV	0.0994	47.06667	6.74471	0.4935	67	27.587	VV	0.1724	120.19996	9.58078	1.2604
29	14.014	VV	0.2095	279.42154	19.69484	2.9300	68	28.245	VV	0.2713	116.07616	5.61247	1.2172
30	14.342	VV	0.1064	43.98084	5.93851	0.4612	69	28.507	VP	0.2112	46.15210	2.90675	0.4840
31	14.486	VV	0.1774	79.43365	6.04171	0.8329							
32	14.767	VV	0.1912	99.47188	7.10946	1.0431							
33	15.050	VB	0.1878	128.43883	9.03874	1.3468							
34	15.432	BV	0.1844	47.37115	3.36102	0.4967							
35	15.610	VV	0.1870	45.43488	3.25157	0.4764							
36	15.888	VV	0.0878	19.28909	3.22361	0.2023							
37	16.093	VV	0.1446	61.74771	5.69234	0.6475							
38	16.290	VV	0.1424	40.06229	4.31005	0.4201							
39	16.458	VV	0.0956	53.86622	8.09390	0.5648							

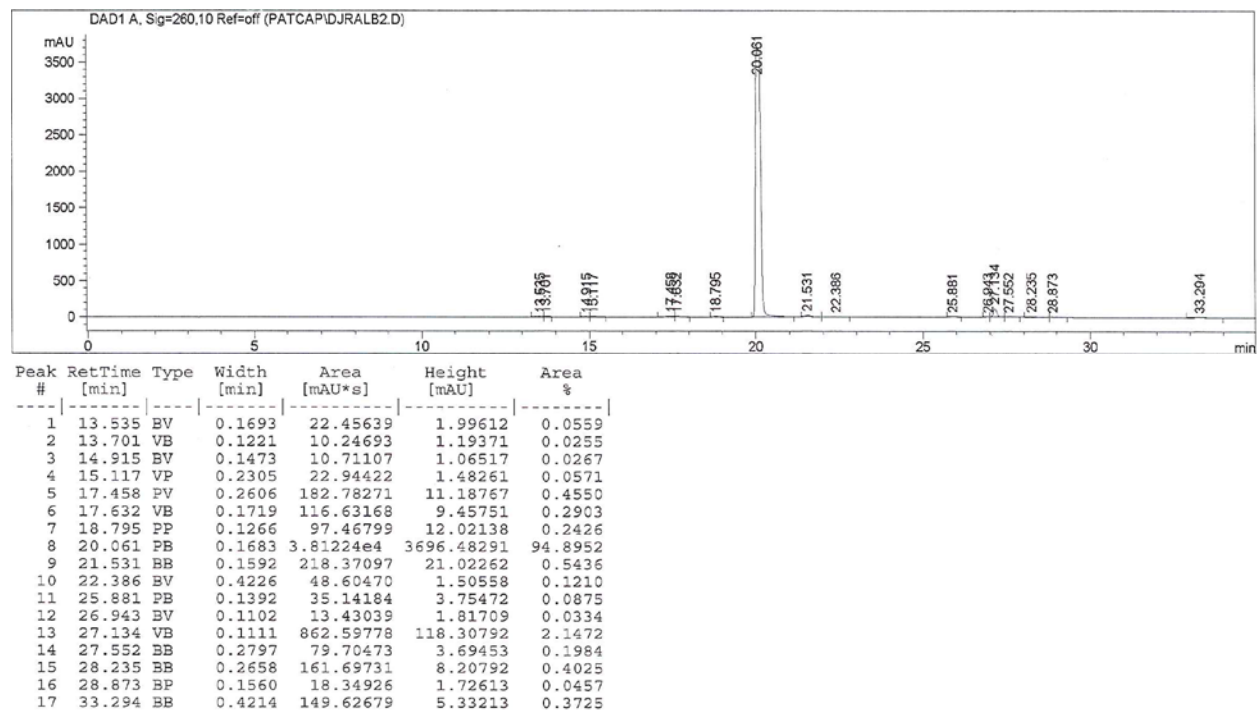


## Black Cohosh: 3

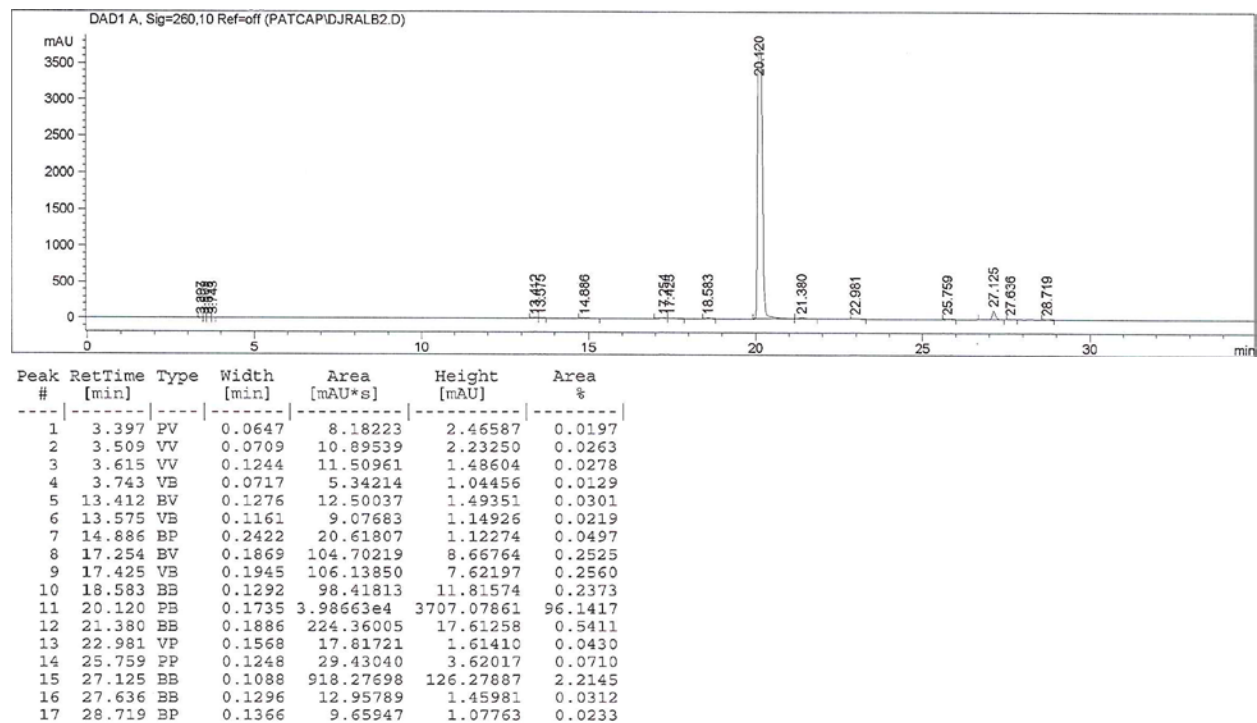


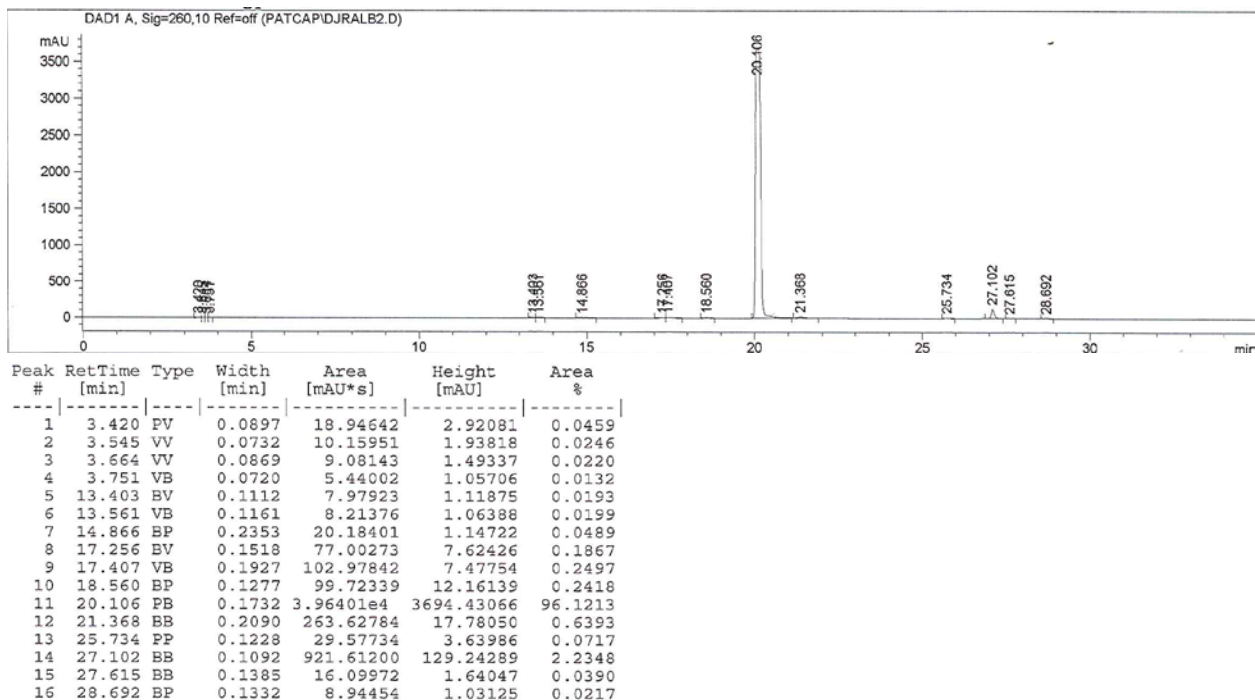
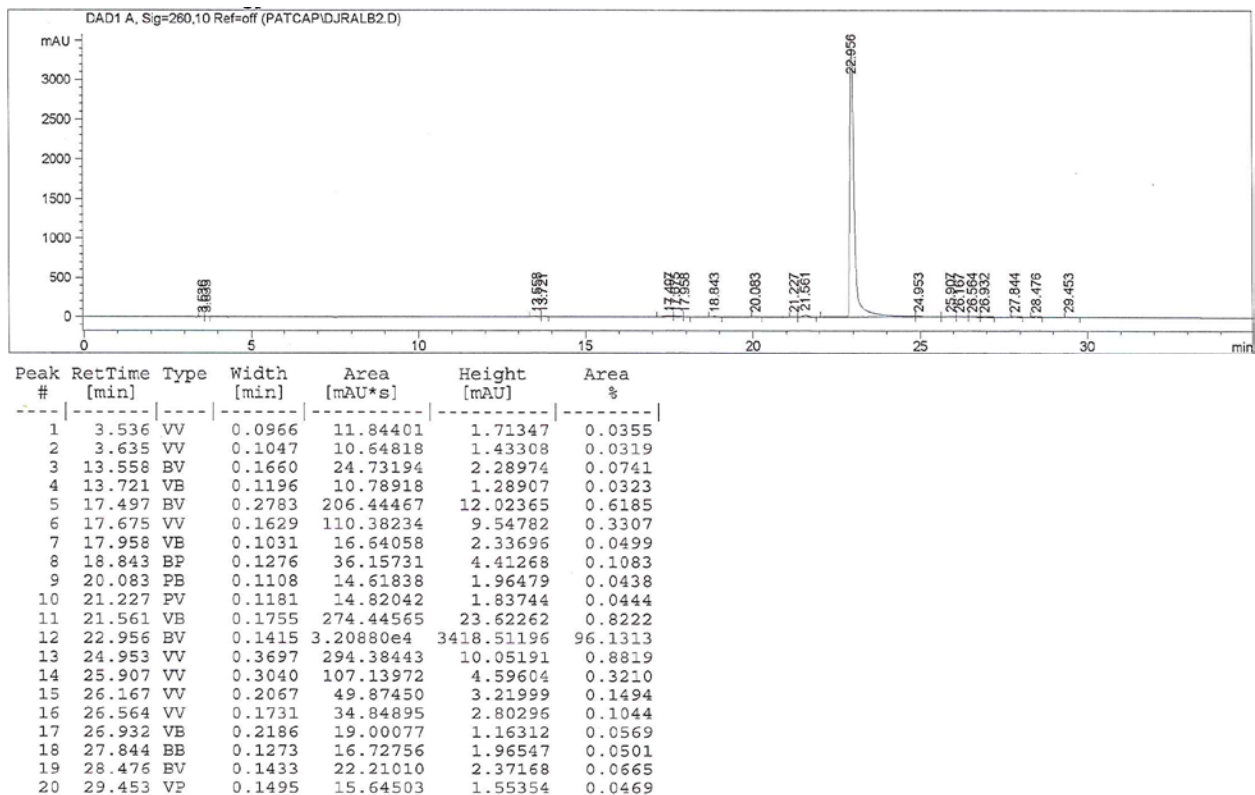


## Daidzein: 1

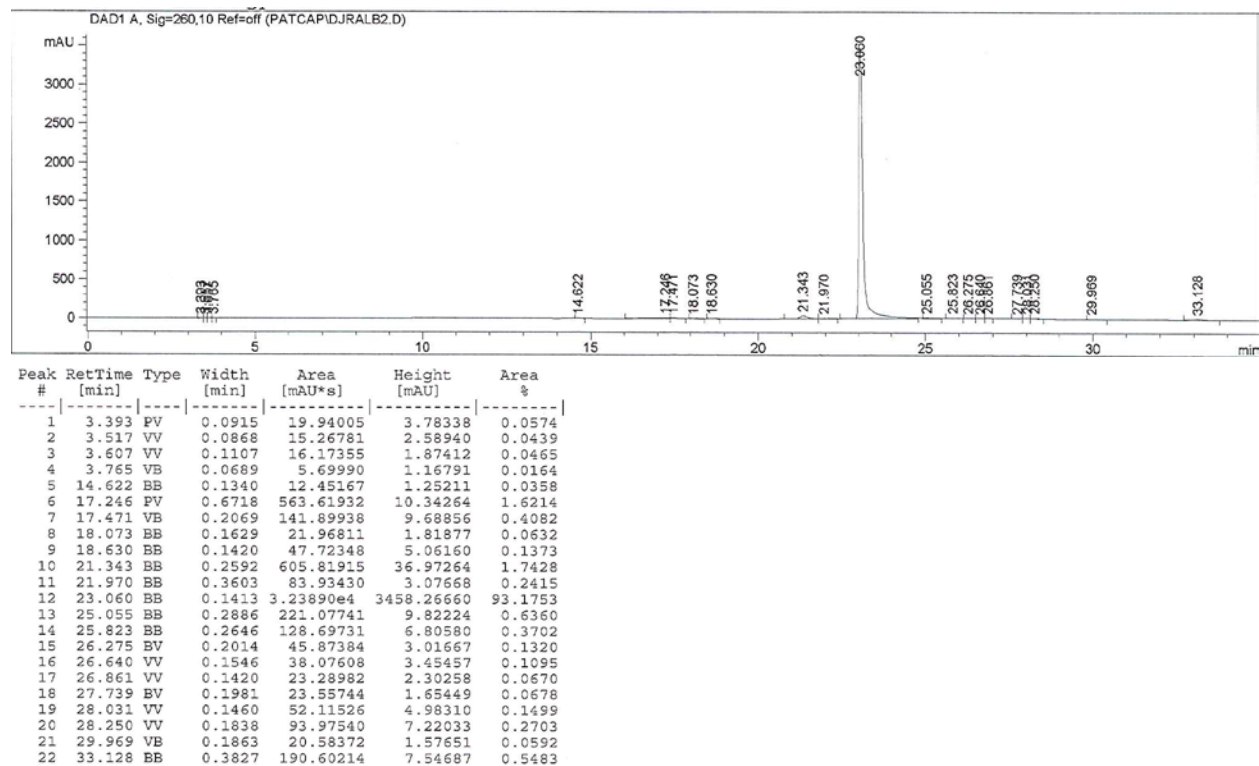


## Daidzein: 2

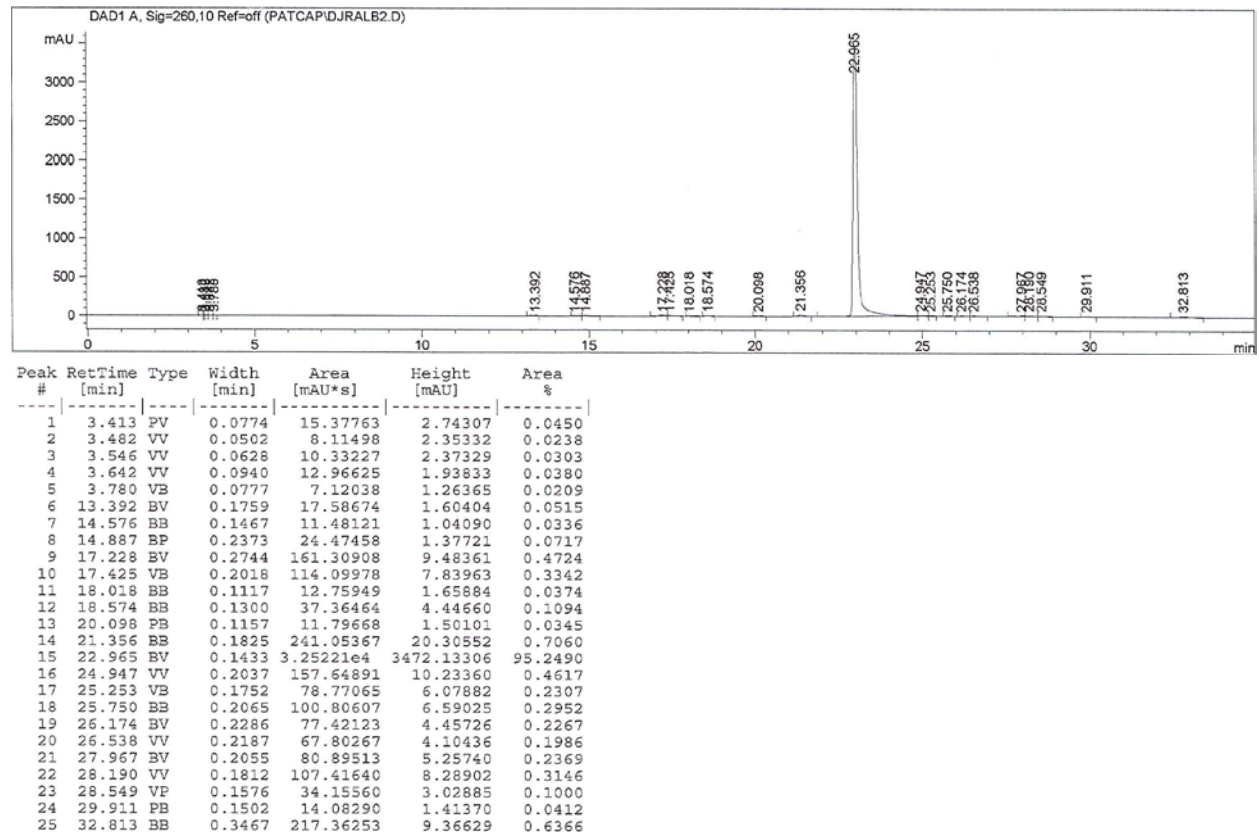


**Daidzein: 3****Genistein: 1**

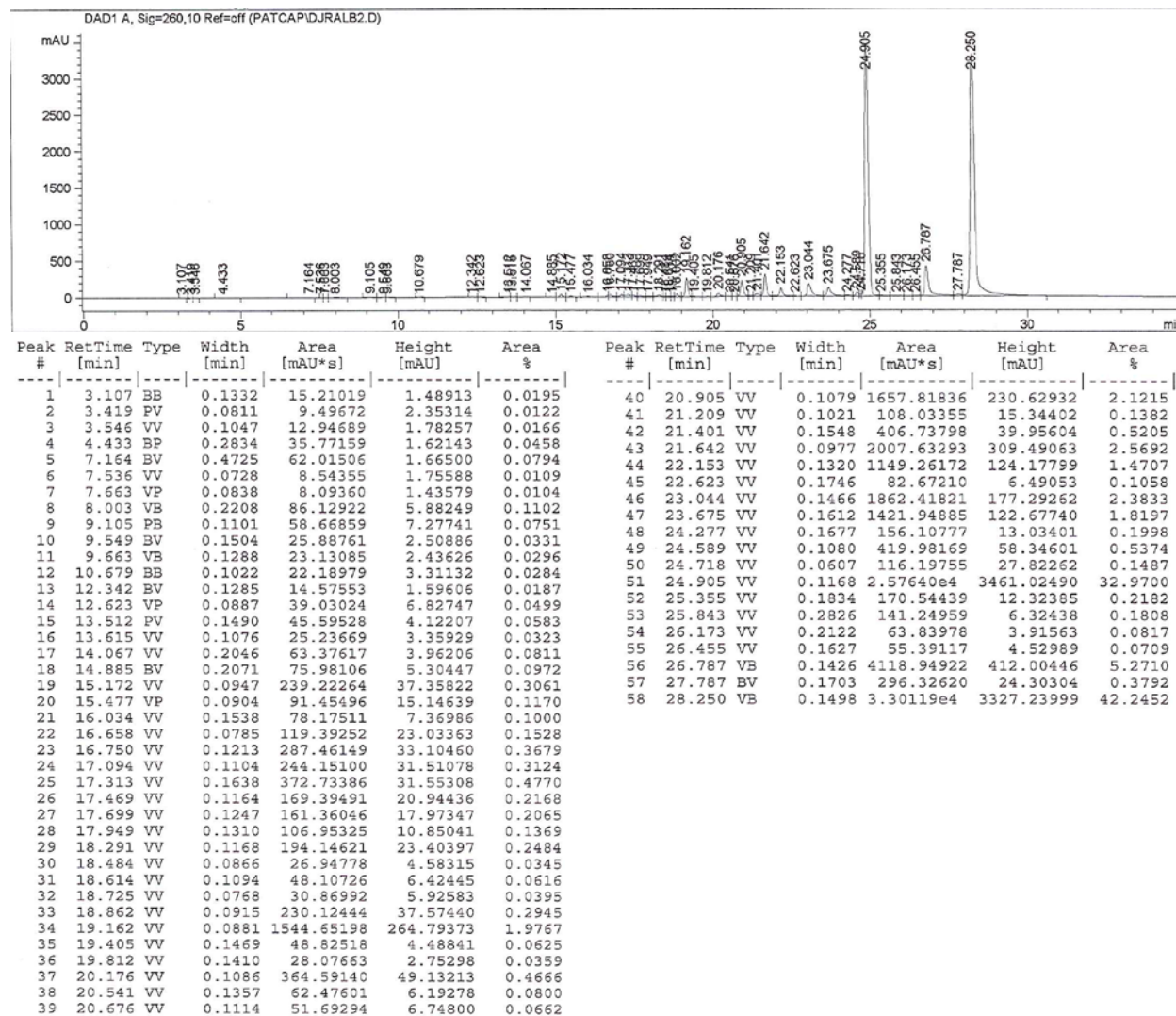
## Genistein: 2



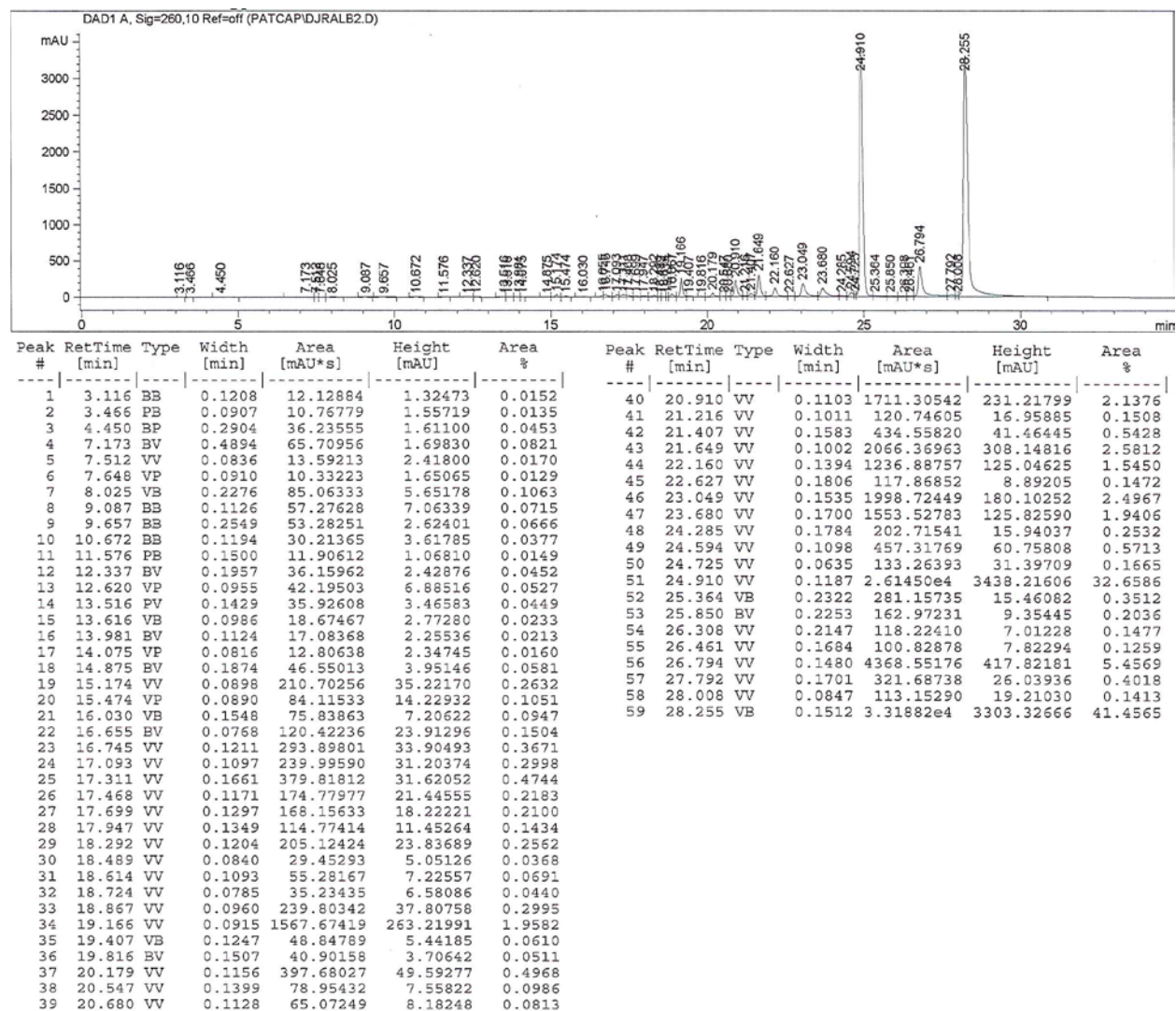
## Genistein: 3



## Promensil: 1

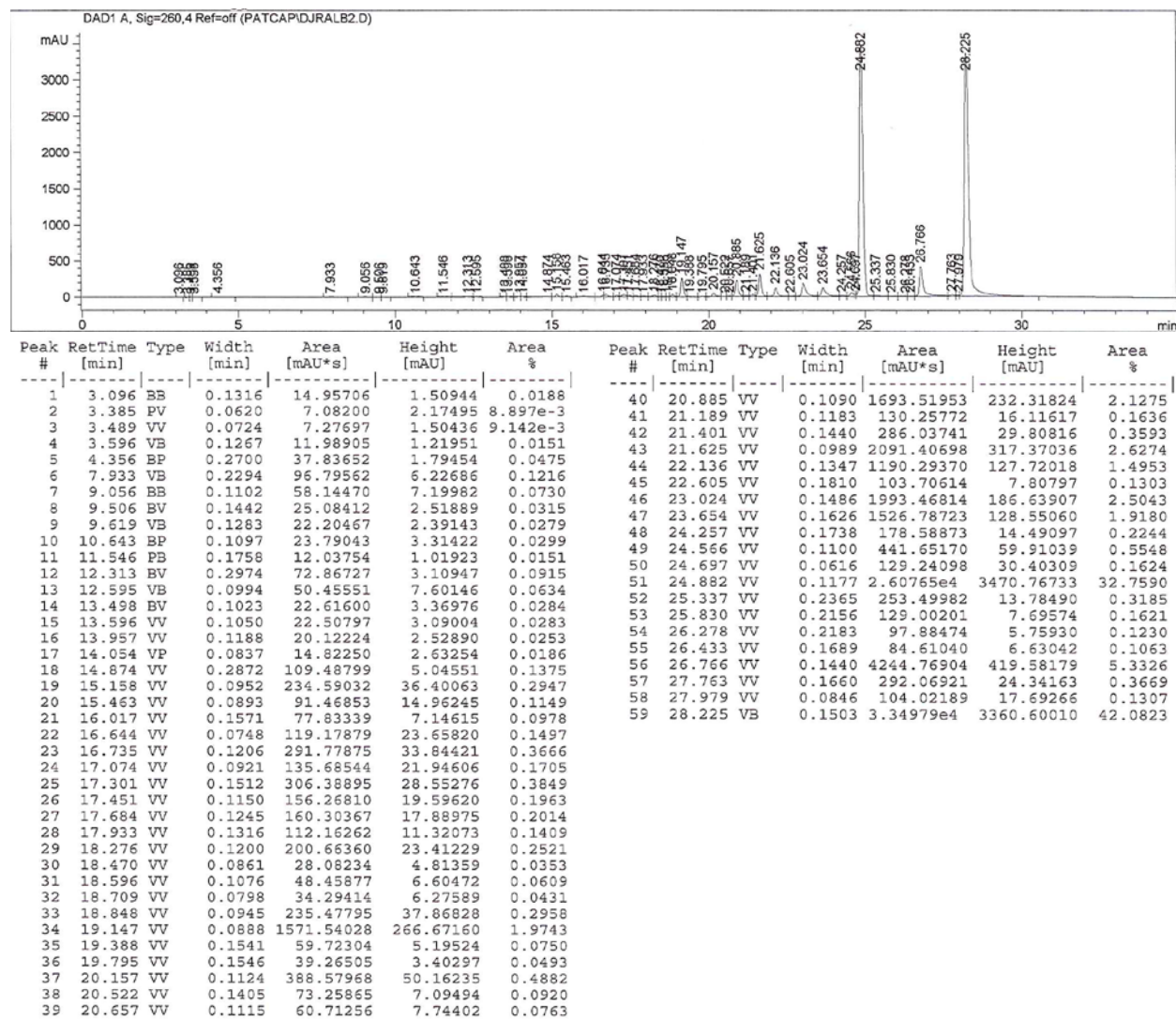


## Promensil: 2

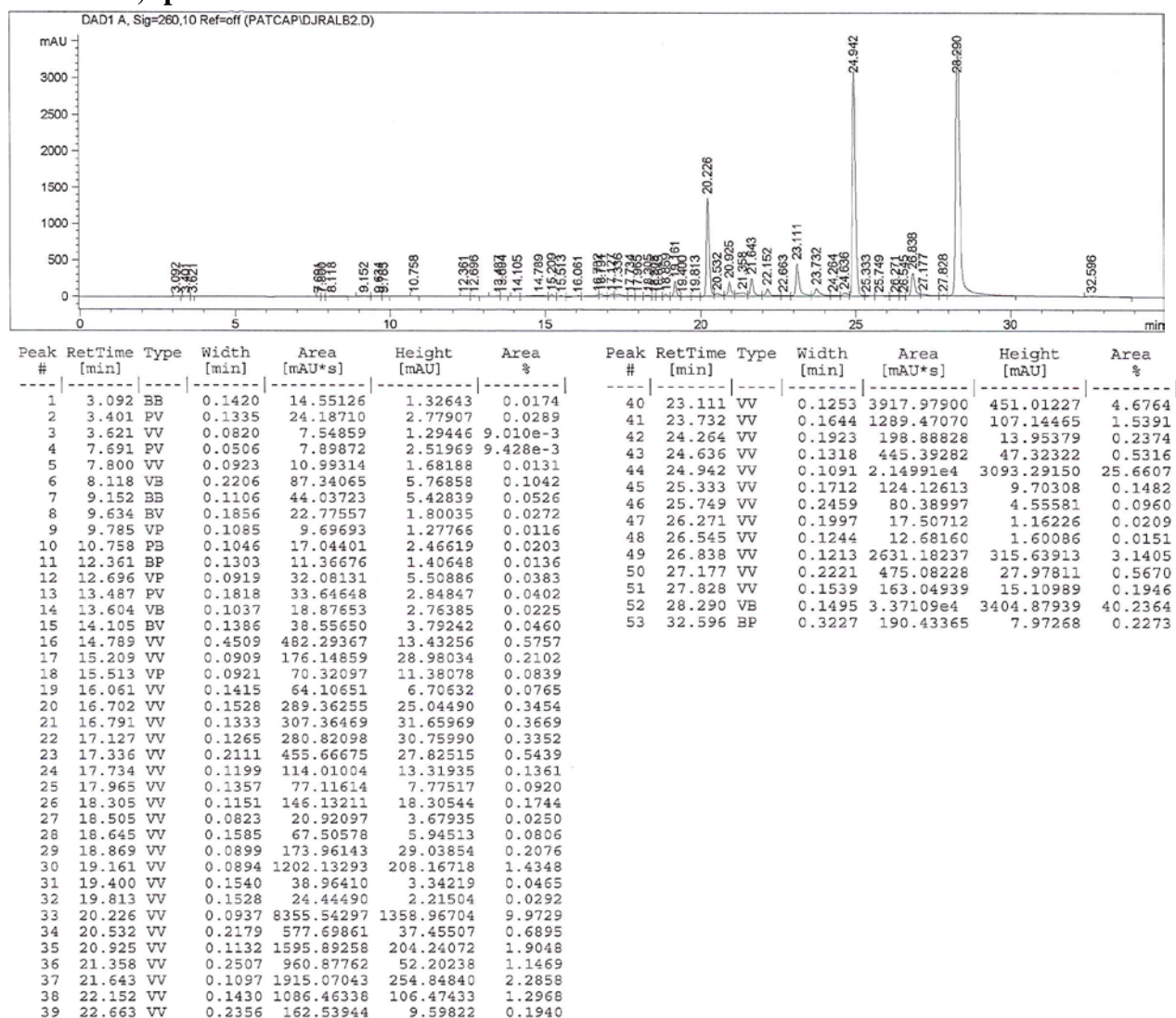




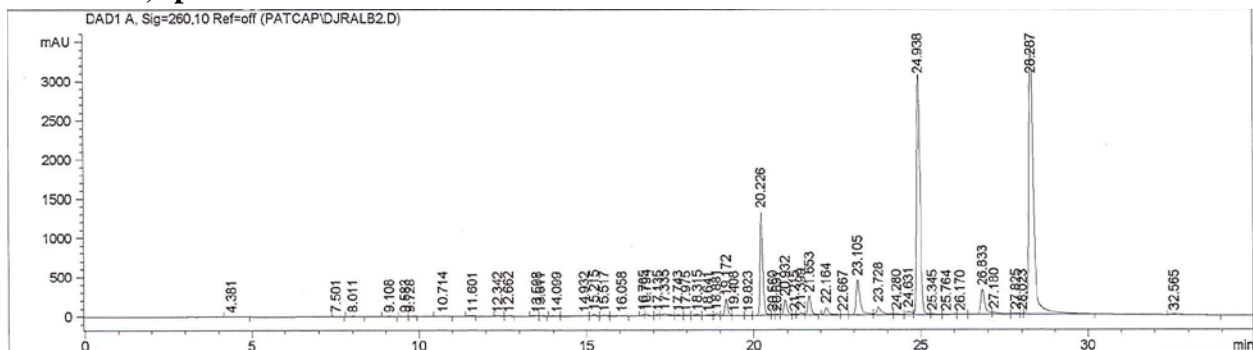
## Promensil: 3



## Promensil, spiked: 1



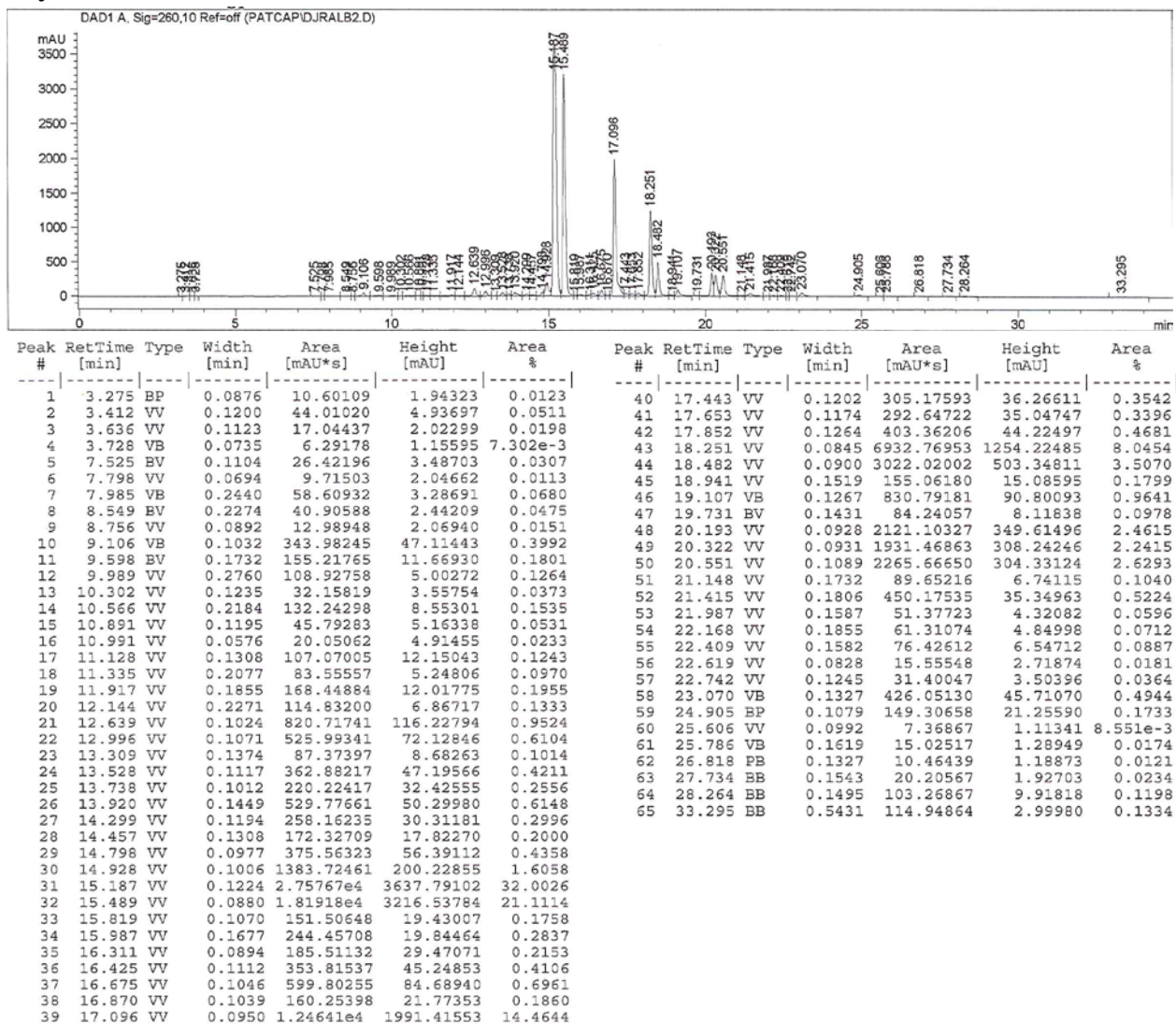
## Promensil, spiked: 2



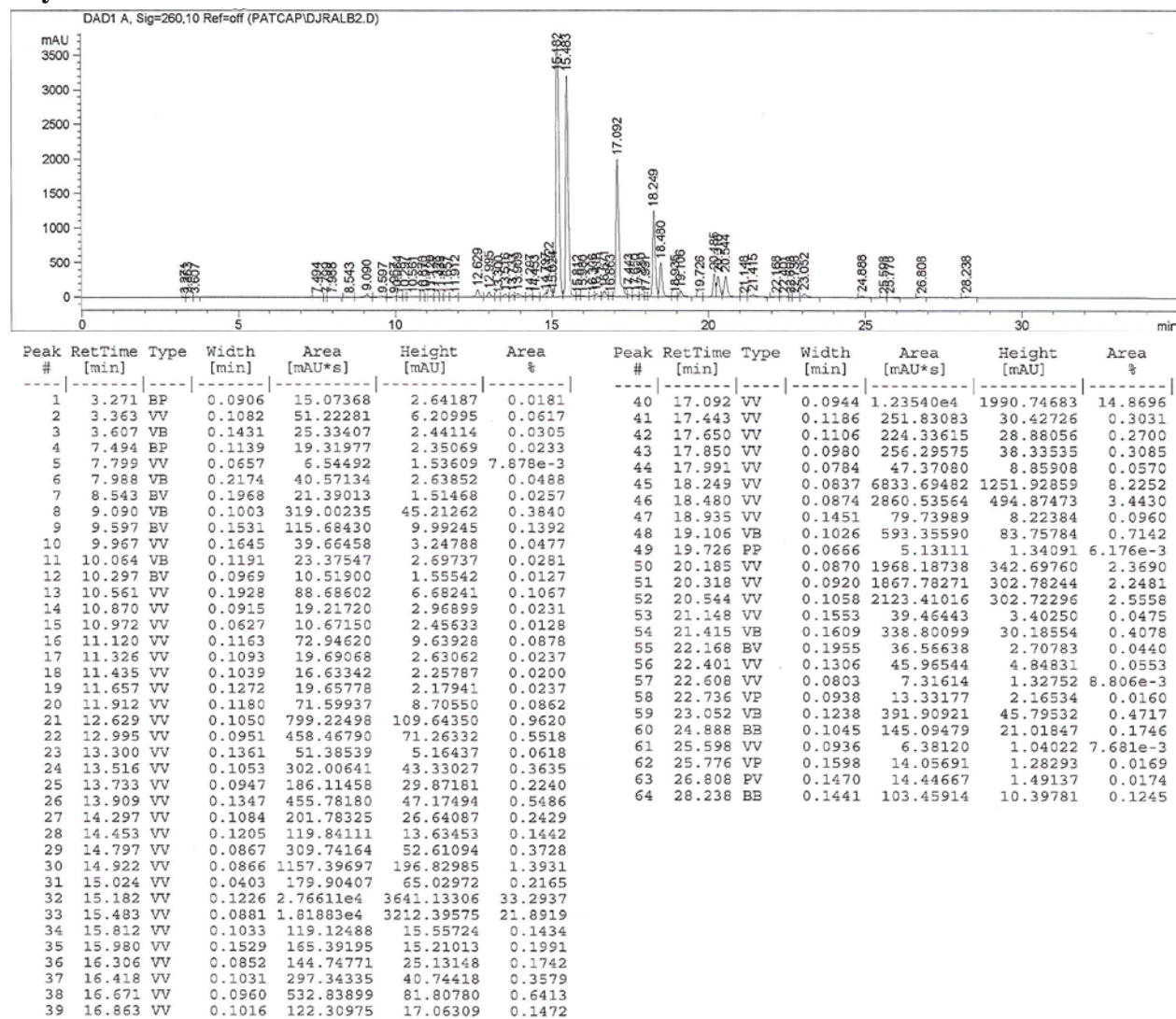
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %	Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.381	BP	0.2815	31.89273	1.45619	0.0393	40	23.728	VV	0.1598	1235.50342	106.14156	1.5222
2	7.501	PP	0.1217	18.57153	2.17372	0.0229	41	24.280	VV	0.1901	162.75635	11.85972	0.2005
3	8.011	PB	0.2185	69.19765	4.73565	0.0853	42	24.631	VV	0.1342	452.54871	47.06530	0.5576
4	9.108	BB	0.1140	44.29646	5.27476	0.0546	43	24.938	VV	0.1080	2.15894e4	3072.10864	26.6000
5	9.583	BV	0.1440	17.04916	1.77561	0.0210	44	25.345	VV	0.1906	149.64960	10.35235	0.1844
6	9.728	VB	0.1151	11.49270	1.38017	0.0142	45	25.764	VV	0.2843	112.05563	5.10074	0.1381
7	10.714	PB	0.1204	20.27864	2.45492	0.0250	46	26.170	VV	0.2164	51.98772	3.08860	0.0641
8	11.601	BV	0.1512	10.20790	1.03417	0.0126	47	26.833	VV	0.1267	2772.75220	321.29565	3.4163
9	12.342	BV	0.1851	36.81514	2.88015	0.0454	48	27.180	VV	0.2273	492.30716	28.24850	0.6066
10	12.662	VB	0.1006	37.66197	5.59236	0.0464	49	27.825	VV	0.1594	187.02760	16.35503	0.2304
11	13.508	BV	0.1241	21.97399	2.72540	0.0271	50	28.023	VV	0.0870	63.90716	10.80978	0.0787
12	13.611	VB	0.1100	20.98957	2.78148	0.0259	51	28.287	VB	0.1494	3.38123e4	3419.79248	41.6596
13	14.099	BV	0.1565	21.79259	1.86229	0.0269	52	32.565	BP	0.3503	211.50014	8.00249	0.2606
14	14.932	VV	0.3048	114.00523	4.87526	0.1405							
15	15.215	VV	0.0946	170.27249	27.37191	0.2098							
16	15.517	VP	0.0974	71.62874	11.08846	0.0883							
17	16.058	VV	0.1509	55.96370	5.58403	0.0690							
18	16.705	BV	0.0736	92.68378	19.45888	0.1142							
19	16.794	VV	0.1175	220.19652	26.35431	0.2713							
20	17.135	VV	0.0975	133.34773	20.06687	0.1643							
21	17.335	VV	0.2192	424.96027	24.88615	0.5236							
22	17.743	VV	0.1251	118.76706	13.43375	0.1463							
23	17.975	VV	0.1330	81.31571	8.25227	0.1002							
24	18.315	VV	0.1167	147.75781	18.19418	0.1821							
25	18.641	VV	0.1943	87.39761	5.98584	0.1077							
26	18.881	VV	0.0928	175.59468	28.94309	0.2163							
27	19.172	VV	0.0907	1213.78601	206.22263	1.4955							
28	19.408	VB	0.1290	29.67384	3.17714	0.0366							
29	19.823	BV	0.1183	12.17225	1.50552	0.0150							
30	20.226	VV	0.0958	8122.50635	1318.90942	10.0076							
31	20.560	VV	0.0905	52.26101	8.18073	0.0644							
32	20.691	VV	0.1150	64.23051	7.89114	0.0791							
33	20.932	VV	0.1031	1328.85425	191.21706	1.6373							
34	21.215	VV	0.0981	85.86880	12.50331	0.1058							
35	21.399	VV	0.1574	318.94138	30.64200	0.3930							
36	21.653	VB	0.0998	1615.87122	242.31406	1.9909							
37	22.164	BV	0.1264	849.09381	98.64030	1.0462							
38	22.667	VV	0.1640	60.48807	4.96990	0.0745							
39	23.105	VV	0.1263	3859.73804	449.17841	4.7555							



## Soy: 1



## Soy: 2



## Soy: 3

